



Science Arts & Métiers (SAM)

is an open access repository that collects the work of Arts et Métiers Institute of Technology researchers and makes it freely available over the web where possible.

This is an author-deposited version published in: <https://sam.ensam.eu>
Handle ID: <http://hdl.handle.net/10985/8097>

To cite this version :

Sandra DOMENEK, Abderrahim LOUAIFI, Alain GUINAULT, Stéphanie BAUMBERGER -
Potential of Lignins as Antioxidant Additive in Active Biodegradable Packaging Materials - Journal
of Polymers and the Environment - Vol. 21, n°3, p.692-701 - 2013

Any correspondence concerning this service should be sent to the repository

Administrator : scienceouverte@ensam.eu



Potential of Lignins as Antioxidant Additive in Active Biodegradable Packaging Materials

Sandra Domenek · Abderrahim Louaifi ·
Alain Guinault · Stéphanie Baumberger

Abstract Due to their polyphenolic structure lignins bear a number of interesting functional properties, such as antioxidant activity. Natural antioxidants are very much looked for in the aim of protection of light or oxygen sensitive goods and are being used in active packaging. Poly(lactide) (PLA)-lignin films were prepared by twin screw extrusion followed by thermo-compression using two different commercial sources of alkali lignins obtained from gramineous plants. A good dispersion of lignin in the matrix was observed. Mechanical properties of the compounded material were merely diminished and oxygen barrier properties slightly enhanced. The chromatographic study of the lignins revealed that the low molecular weight fraction of both lignins increased during the polymer processing. The migration of low molecular weight compounds in an ethanol/water solution simulating fatty foodstuff was performed and the antioxidant activity of the extract was analysed. It was found that the activity increases with increasing severity of the heat treatment because of the generation of free phenolic monomers

during processing. These results open an interesting way for application of lignins as an active compound in packaging materials. Lignins do not impair the mechanical and barrier performance of the polymer and the plastics processing even allows for the generation of active substances.

Keywords Antioxidant · Food packaging · Lignin · Biodegradable polymer · Poly(lactic acid)

Introduction

Lignin is the second major component of wood and annual plants [1] after cellulose. Derived from native lignin, industrial or technical lignins are available in large amounts as byproduct of the production of cellulose pulp in papermaking. Their quantity is supposed to keep increasing with the development of the lignocellulosic biorefinery and the production of second generation bioethanol. These by-products contain highly branched, irregular phenolic macromolecules, the structure of which is depending on botanic origin, harvesting period and extraction process. Industrial lignins are mainly used for energy production, but with regards to the huge amount available, valorization of a part of this abundant feedstock in high value applications is certainly attracting and also going to play a role in the economic feasibility of the production of bioethanol from lignocellulosic feedstocks [2, 3].

The blending of lignin with polymers has already been demonstrated [4]. Due to their aromatic structure and the occurrence of phenol residues, lignins have multiple functionalities, being able to act as compatibilizers [5], plasticizers [6], hydrophobizing agents [7, 8], flame retardants [9] or optical modifiers [10] or stabilizers [11]. Furthermore, properties potentially interesting in high value applications,

S. Domenek · A. Louaifi
AgroParisTech, UMR 1145, 91300 Massy, France

S. Domenek (✉)
1, rue des Olympiades, 91744 Massy Cedex, France
e-mail: sandra.domenek@agroparistech.fr

S. Domenek
INRA, UMR 1145, 91300 Massy, France

A. Guinault
CNAM, PIMM- P2AM, 75141 Paris, France

S. Baumberger
Institut Jean-Pierre Bourgin, UMR 1318 AgroParisTech-INRA,
Pôle PAVE, 78000 Versailles, France

such as antimicrobial activity [12] or cytotoxic effects [13] have been shown. The antioxidant properties of lignins stem from their radical scavenging capacity [13–17], following the general mechanism of monomeric phenols. The hindered phenolic hydroxyl groups being part of the lignin structure act as proton donors and are able to stabilize the radical in the quinone resonance structure [16]. Radical scavenging activity is described for a large number of lignin sources, ranging from black liquors [18] to the description of different botanic sources such as miscanthus [19], oak chips [20], bamboo [21] and many others. In general, radical scavenging activity of lignin is negatively influenced by increasing heterogeneity, dispersity, molecular weight average and carbohydrate admixtures [13, 15, 16]. Lignins extracted in sodium hydroxide aqueous medium (alkali lignins) have been shown to have higher activity than other types of lignins, such as lignosulfonates, Kraft, or steam explosion lignin, with a radical scavenging activity similar to that of epicatechin, used as a reference antioxidant [14, 17]. Alkali lignins contain a fraction of free phenolic monomers, mainly ferulic and p-coumaric acids [8], likely to account for this high radical scavenging activity.

In the field of material applications, the antioxidant activity has been investigated in the aim of using lignin as a natural substitute of synthetic antioxidants required for the formulation of polyolefins [22–25]. Pouteau et al. [26] showed that the compatibility between lignin and polymer matrix could be successfully predicted using the solubility parameters calculated after the Small, Van Krevelen and Hoy method.

The same mechanism protecting polyolefins against radical oxidative degradation is valuable for protection of fatty acids. The use of natural antioxidants to protect lipids and food has already long history [27–30]. For example, Ugartondo et al. [31] showed the capacity of lignin to prevent lipid peroxidation in human red blood cells. Polymer packaging materials are interacting with packed goods due to mass transfer between both partners, which is influencing food quality [32, 33]. It is interesting to make positive use of such an interaction to ensure a sustained delivery of an active substance such as the antioxidant into the food stuff. In that aim, active packaging is now increasingly used as a reservoir for active substance delivery [34–37], complementing other techniques such as modified atmosphere packaging [38], edible coating [39, 40] or oxygen scavengers [41, 42].

The objective of the present study is to investigate the potential of lignin to be used as a natural antioxidant additive in an active food packaging. The packaging matrix chosen was poly(lactide) (PLA) because of its renewable character. The inclusion of natural antioxidants in PLA in the aim of developing an active material has already been suggested in literature [43–45]. Furthermore, some studies already exist pointing to the feasibility of inclusion of lignin into PLA [5, 9, 46, 47]. Two commercial alkali lignins were chosen

because of their expected radical scavenging activity and their difference in the free phenolic monomers content. PLA was blended with lignin by twin screw extrusion and film samples were fabricated by thermo-compression. The effect of lignin quantity and processing on the radical scavenging activity was assessed in an ethanol/water solution being a stimulant of fatty foods recommended by the European regulation for food contact materials [48].

Materials and Methods

Poly(lactide), PLA 2002D, was purchased from NatureWorks (USA). Lignin PROTOBIND 1000, extracted by an alkali process from a mix of wheat straw (*Triticum* sp.) and sugar cane (*Saccharum munja*), and lignin SARKANDA, extracted by the same process from sugar cane (*Saccharum munja*), were obtained from GRANIT SA (Switzerland). Lignin PROTOBIND 1000 and SARKANDA differed in their content in free phenolic monomers: 8.0 and 0.5 mg g⁻¹, respectively [8].

Sample Preparation

PLA was dried at 80 °C for 8 h in a SOMOS dryer (Mann & Hummel ProTech, Germany). Final water content was around 720 ppm. Lignin powder was dried in an oven under vacuum at 60 °C for 15 h. A dry blend of PLA pellets and lignin of desired quantity was realized and introduced into the feeder of a twin-screw extruder (Thermo-Haake Rheomix PTW 16/40, Germany). Barrel temperature was 180 °C and die temperature 170 °C, rotating speed was set to 400 rpm. Throughput was fixed 1,000 g h⁻¹. The extrudate was cooled in a water bath to room temperature and pelletized with the help of a Scamia (France) pelletizer. Pellets were then dried at 80 °C for 8 h under vacuum and stored over silica gel. Sample films were fabricated by thermo-compression under a heating press (Télémechanique 15T N1, France) sandwiched between two Teflon sheets and two steel plates. Samples were melted for 2 min at 185 °C without pressure, and then pressure was applied in three times to eliminate air bubbles: 30 s at 30 bar, 30 s at 50 bar and 1 min at 150 bar. In order to vary the heat treatment of the samples, the films were maintained in the heating press at 185 °C under a pressure of 50 bar for different times. Films thickness, which was measured with a hand-held micrometer, lay between 200 and 300 µm. The PLA reference film was prepared only by thermo-compression.

Material Characterization

Lignin/PLA blend morphology was observed with the help of an transmission optical microscope (Nachet, France) on

samples of 10 μm thickness prepared by a microtome (Leica RM 2255, France).

Glass transition temperature (T_g) and crystallinity degree (χ_c) were analyzed by differential scanning calorimetry (DSC). The DSC apparatus (Q100, TA Instruments, France) was calibrated with the help of indium and zinc standards before analysis. Samples (10 mg) were loaded into hermetic aluminum crucibles (TZero) and a heat-cool-heat cycle was performed between 0 and 200 $^{\circ}\text{C}$ with a heating and cooling rate of 10 $^{\circ}\text{C min}^{-1}$. The enthalpy of PLA cold crystallization and melting was measured at the first heating run. The crystallinity degree (χ_c) was calculated with Eq. 1, where ΔH_m is the enthalpy of melting, ΔH_{cc} is the enthalpy of cold crystallization and ΔH_m^0 is the enthalpy of fusion per mol of repeating unit of the perfect crystal of infinite size, being 93 J g^{-1} [49]:

$$\chi_c = \frac{\Delta H_m - \Delta H_{cc}}{\Delta H_m^0} \quad (1)$$

The T_g was measured in the second heating run on the midpoint of the heat capacity step. Analyses were carried out in triplicate.

The uniaxial tensile testing was carried out at room temperature and at 5 mm min^{-1} with an Instron tensile testing machine (Instron Model 4507) equipped with pneumatic jaws on type I BA dumbbell shaped samples (in accordance with the EN ISO 527 standard). Each value is an average of 5 measurements.

The oxygen transmission rate (OTR) was monitored at 23 $^{\circ}\text{C}$, 0 % RH, and a pressure gradient of 1 atm with a Systech 8001 apparatus. Oxygen permeability was then obtained by dividing the OTR by the film thickness. Film thickness was determined as the average of 9 thickness measurements on the film sample.

Chromatography

Molecular weight distribution of PLA, lignins and blends was determined by size exclusion chromatography (SEC). SEC was performed on a PL-gel mixed C column (Polymer Laboratories, 5 μm , 600 \times 7.5 mm) using stabilized THF (Carlo Erba SDS, HPLC quality) as eluent at a flow rate of 1 mL min^{-1} . Detection was done by a diode array detector (Dionex Ultimate 3000, France) in series with a refractive index detector (Shodex RI 101, France). The lignin fraction was detected at 280 nm wavelength by the diode array detector and PLA by the refractive index detector. Injection volume was 10 μL . Calibration was performed with monodisperse polystyrene standards (EasyCal, Polymer Laboratories) with molecular weight ranging from 580 to 210,500 g mol^{-1} . Lignins and PLA-lignin blends were acetylated using a 1:2 pyridine/acetic anhydride mixture (v/v, 48 h at room temperature) and recovered according to

Gellerstedt [50] prior to SEC analysis. Solutions of acetylated lignins in tetrahydrofuran (about 10 mg mL^{-1}) were ultrafiltrated on a 0.45 μm PTFE filter (Millipore) before injection.

The identification and quantification of low molecular weight phenolic compounds migrating in an ethanol/water (95/5 v/v) solution was run on a polydimethylsiloxane capillary column (DB1, Supelco, 30 \times 0.25 m, 0.25 μm) GC/MS (Saturne 2100, Varian) with an ion trap spectrometer detector (IE 70 eV, positive mode) under helium (pressure 7 psi, 1 mL min^{-1} flow rate). Injector temperature was 270 $^{\circ}\text{C}$. The temperature program was the following: a ramp from 40 to 110 $^{\circ}\text{C}$ at 30 $^{\circ}\text{C min}^{-1}$, then from 110 to 200 $^{\circ}\text{C}$ at 2 $^{\circ}\text{C min}^{-1}$ and finally from 200 to 260 $^{\circ}\text{C}$ at 3 $^{\circ}\text{C min}^{-1}$. Sample preparation comprised a migration step from PLA and silylation. Migration was performed at room temperature for 48 h by immersion of 4 cm^2 of film in 12 mL ethanol/water (95/5 v/v [48]) under stirring. After migration, the solvent was evaporated and the dry sample was weighted. Then it was adjusted in 100 μL dichloromethane (Carlo Erba SDS) containing 0.495 mg mL^{-1} n-nonadecane (C19) and n-henicosane (C21) alkane standards (Sigma Aldrich). The solution was dried over Na_2SO_4 (Sigma Aldrich). An aliquot of 10 μL was silylated by addition of 100 μL trimethylsilyl(1E)-2,2,2-trifluoro-N-(trimethylsilyl)ethanimidoate (BSTFA, Aldrich) and 10 μL pyridine (Merck). The reference solution was prepared in the same way and contained the following commercial phenols with a concentration of 1 mg L^{-1} : vanillin (Fluka, purum), syringaldehyde (Aldrich Chimie, purity 98 %), vanillic acid (Fluka, purum), syringic acid (Fluka, purum), *p*-coumaric acid (Sigma Aldrich) and ferulic acid (Fluka, purum). Semi-quantitative analysis was performed with response factors of the internal standards C21 and C19. Migrations were carried out in duplicate. Experimental error was ± 20 %.

Antiradical Activity

The antiradical activities of the films were probed using a spectroscopic method based on the disappearance of the absorption band at 515 nm of the free radical, 2,2-diphenyl-1-picrylhydrazyl (DPPH $^{\circ}$) (Sigma-Aldrich, France) upon reduction by an antiradical compound [51]. The test consisted in adding 77 μL of the antioxidant solution into 3 mL of a DPPH $^{\circ}$ solution in methanol (6×10^{-5} mol L^{-1}) and following the intensity of the 515 nm absorption band over time (spectrophotometer Uvikon 810 Kontron, France). For the test of film samples, pieces of 1 cm^2 were directly immersed into 3 mL of the methanolic DPPH $^{\circ}$ solution. For the test of antiradical activity of migrating substances, 1 cm^2 of film was immersed in 3 mL methanol or in 3 mL ethanol/water (95/5 v/v) for 24 h at ambient temperature.

The ethanol/water mixture was used as simulant for fatty food, as suggested by the European Regulation, whereas methanol was used as widespread extraction solvent for phenolic monomers. The supernatant was recovered after 24 h and evaporated to dryness before being adjusted in 77 μL methanol for the test. Experimental error of the determination of the remaining quantity of DPPH $^\circ$ was $\pm 7\%$. Ethanol (p.a. quality) and methanol (p.a. quality) were purchased by Carlo Erba.

Results and Discussion

Composite Material Characteristics

The material properties of the lignin/PLA blends were analyzed, in order to assure the quality of the packaging film. The two step process used in this study (twin screw extrusion and thermocompression) yielded a homogeneous dispersion of lignin particles of around 5–20 μm diameter inside the PLA matrix, which is shown in Fig. 1 for the example of lignin PROTOBIND 1000. The molecular weight distribution of PLA in the blended films and their mechanical, thermal and oxygen barrier properties are shown in Table 1. It can be observed on the neat sample that the sample fabrication process brought about a decrease in the molecular weight average (M_w) of PLA. Such a decrease is often observed as PLA is very sensitive to thermohydrolysis caused by remaining traces of water [52]. The inclusion of lignin into PLA had no further effect on its M_w . The glass transition temperature (T_g) of the lignin/PLA films was slightly lower than the one of the neat PLA, but no significant differences between the blends were detected. This may be attributed to a plasticizing effect. Indeed, a decrease of the T_g of PLA due to the introduction of the natural antioxidant resveratrol was also observed by Hwang et al. [45] and explained by plasticizing. Moreover, aromatic molecules such as benzaldehyde have already shown their ability to decrease the T_g of PLA [53]. The low molecular weight fraction of lignin has therefore probably a plasticizing effect. PLA/lignin films were amorphous under the given process conditions, no degree of crystallinity differences were observed and so no nucleating effect was observed. Results were the same for both lignin grades used.

As shown in Table 1, the inclusion of more than approximately 1 wt% of lignin decreased the yield stress, but it stays at a relatively high value. The elongation at break decreased slightly for the PLA/lignin SARKANDA formulation with the increase of lignin content. However, as PLA on itself is already a brittle material [54–58], further decrease was difficult to detect. The measured strain at break values of neat PLA data were in agreement with the

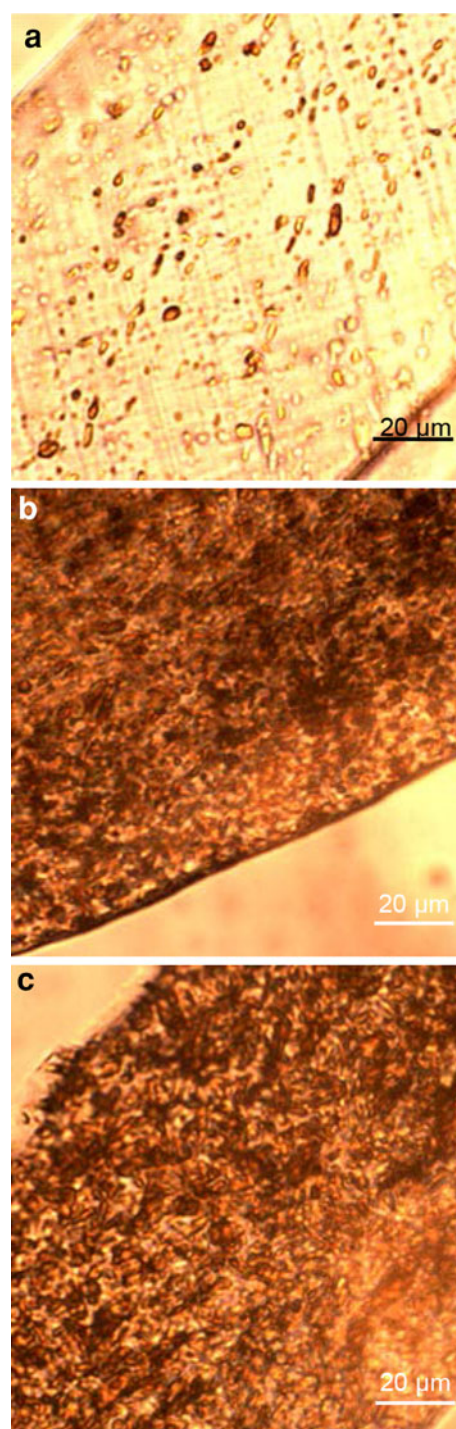


Fig. 1 Lignin distribution inside PLA obtained by twin screw extrusion and thermo-compression of PLA-PROTOBIND 1000 dry blend for 1 (a), 5 (b) and 10 wt% (c). Magnification $\times 50$

data given by the provider in the PLA 2002D datasheet and with literature results [56, 59]. The relatively low decrease of yield stress and strain at break were certainly due to the counterbalance of two effects: the addition of high molecular mass lignin lead to increase of yield strength and

Table 1 Physicochemical characteristics of PLA-lignin blends

	Lignin (wt%)	M_w (g/mol)	I_p	T_g (°C)	χ (%)	σ (MPa)	ε (%)	$P(O_2) \times 10^{18}$ (m ³ m m ⁻² s ⁻¹ Pa ⁻¹)
PLA pellets		209,000	1.18	60 ± 1	37 ± 1			
PLA film		139,000	1.91	61 ± 1	2 ± 1	67 ± 3	7 ± 1	2.3 ± 0.3
PLA/PROTOBIND 1000	1	142,000	1.81	55 ± 1	1 ± 1	55 ± 6	7 ± 3	2.1 ± 0.1
PLA/PROTOBIND 1000	5	151,000	1.64	53 ± 2	1 ± 1	56 ± 3	7 ± 5	2.0 ± 0.1
PLA/PROTOBIND 1000	10	132,000	1.87	51 ± 2	1 ± 1	52 ± 8	5 ± 1	1.8
PLA/SARKANDA	1	126,000	1.74	56 ± 1	1 ± 1	56 ± 8	6 ± 1	2.2 ± 0.1
PLA/SARKANDA	5	144,000	1.58	54 ± 1	1 ± 1	49 ± 2	4 ± 1	2.0 ± 0.2
PLA/SARKANDA	10	135,000	1.71	52 ± 2	1 ± 1	53 ± 2	5 ± 1	1.7 ± 0.1

M_w molecular weight average, I_p dispersity index, T_g glass transition temperature, χ crystallinity degree, σ yield stress, ε strain at break, $P(O_2)$ oxygen permeability

decrease of strain at break, while the plasticization of PLA by low molecular mass lignin lead to decrease of yield strength and increase of strain at break. The effect of lignin on the mechanical properties data was also consistent with Ouyang et al. [46] who observed a decrease in the maximum strength for the blend of PLA/cellulolytic enzyme lignin and a slight decrease of strain at break. Comparable behaviour was also observed for PLA/Vanille HW lignin samples [47]. In conclusion, the blending of PLA with lignin did not worsen the good mechanical performance of the materials. This pointed to compatibility between lignin and PLA, a result also pointed out by Ouyang et al. [46] and Li et al. [47]. This compatibility might be optimized in choosing proper lignin grades, following the suggestion of Pouteau et al. [26].

The oxygen barrier properties of the different materials are also given in Table 1. Neat PLA oxygen barrier properties are in accordance with published data [54, 60–65]. Interestingly, the inclusion of at least 10 wt% of lignin yielded a decrease in oxygen permeability of around 20 %. As PLA crystallinity was not modified by the lignin addition, it would mean that lignin has probably lower oxygen permeability than PLA, and might therefore be able to introduce a barrier effect in the polymer.

In conclusion, the inclusion of both lignins at different weight percentages did not change notably the mechanical properties of PLA and even helped to improve oxygen barrier properties. Therefore, a valuable packaging film for rigid containers might be produced within the limits of application drawn by the intrinsic properties of PLA.

Characteristics of the Lignin Charge

The structural characteristics of the lignins in the PLA matrix were followed during the film fabrication process. The Fig. 2 shows the evolution of molecular weight distribution of the samples with the help of the chromatographic

profile in UV detection. In agreement with Zheng et al. [8] the pure lignin samples (Fig. 2a) show a large molecular weight distribution indicating the presence of three populations: a polymeric fraction with low concentration yielding the tailing of the main peak towards small retention times, an oligomeric fraction present in higher concentration with a maximum about 17 min of retention time and then a monomeric fraction corresponding to the peak at 19.5 min only detected in PROTOBIND 1000. The M_w of lignin PROTOBIND 1000 and SARKANDA was calculated to be of 3,000 and 2,600 g mol⁻¹, respectively, with, for both, a dispersity index of 3. Such high dispersity indices are usually observed in the case of lignins [8, 66]. The monomeric fraction present in PROTOBIND 1000 was previously found to be composed of *p*-coumaric and ferulic acid, acetosyringone, syringaldehyde and vanillin [8]. The superposition of chromatography profiles of the PLA/lignin blends with neat PLA are shown in Fig. 2b, c. PLA UV profile exhibited a peak at 21 min most probably due to the use of additives in the commercial PLA grade. Consequently, this peak was present on all the profiles and hindered the detection of possible low molecular weight compounds present in lignins. However, the retention time of the highest peak of PROTOBIND 1000 lignin shifted to slightly lower values (from 17 to 17.5 min) and the shoulder detected at about 18 min and at 19 min seemed to increase (Fig. 2b), which suggested a decrease in the lignin molecular weight. In the case of SARKANDA lignin (Fig. 2c), the same shift to higher retention times (from 17 to 17.5 min) of the main peak was observed and the appearance of a shoulder in the main peak at approximately 18.5 min retention time could be observed indicating that phenolic monomers were released during the blend processing. This monomeric fraction remained however in lower proportion than for PROTOBIND 1000 (Fig. 2b, c). The M_w corresponding to the main distribution (peak at 21 min excluded) of the PLA-10 wt% PROTOBIND 1000 and SARKANDA

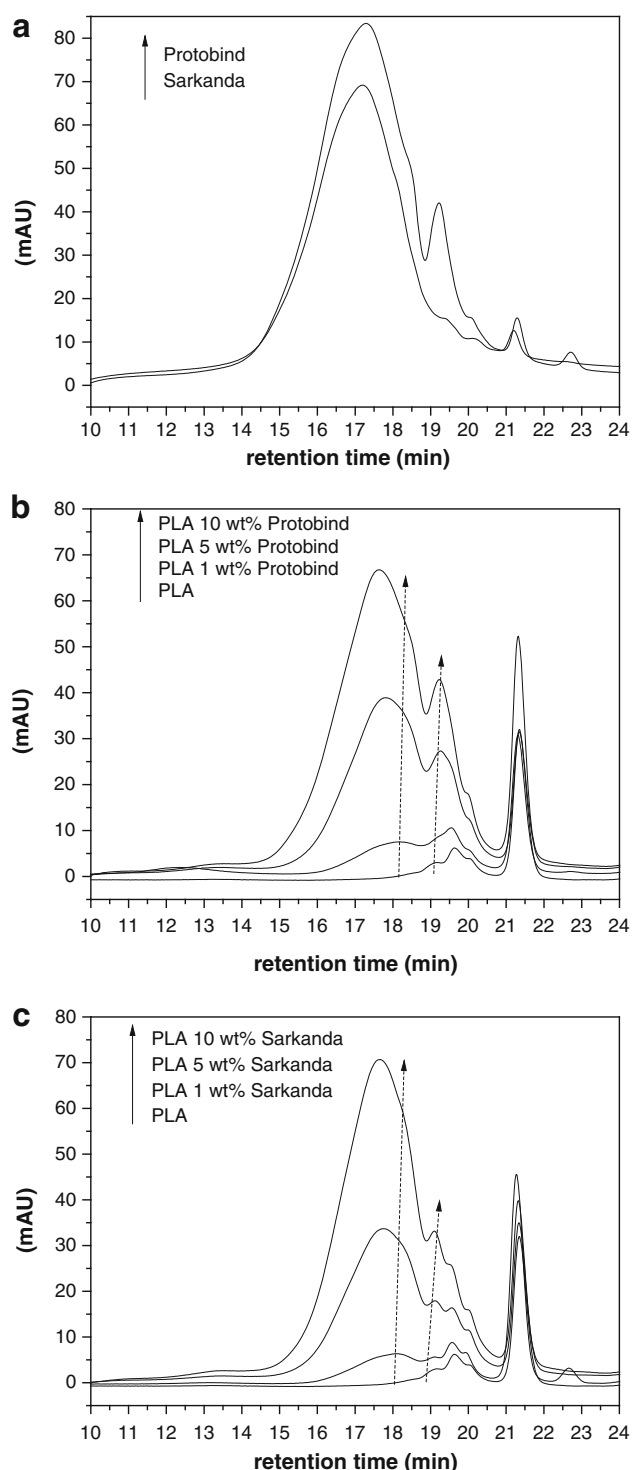


Fig. 2 Size exclusion chromatogram of lignins (a), PLA-PROTOBIND 1000 (b) and PLA-SARKANDA blends (c) fabricated by twin screw extrusion and thermocompression. Curves correspond to the order indicated by the arrow in the legend

blends was calculated to be $1,300 \text{ g mol}^{-1}$, which corresponds to a $\sim 50 \%$ decrease with respect to the initial pure lignin samples.

In conclusion, upon thermal processing for film production, the proportion of low molecular weight compounds increased for both lignin grades, most probably as the consequence of lignin partial depolymerization. These molecules might be the origin of the plasticizing effect observed (Table 1) and discussed before. Furthermore, the low molecular weight fraction being composed of molecules carrying free phenols, the plastics processing might be able to generate additional molecules having an anti-radical activity by the thermal degradation of the lignin charge.

Antiradical Activity of PLA-Lignin Films

Radicals originating from oxygen exist naturally in the atmosphere or can be created by thermal processing or irradiation of packaging and food [67]. Those radicals act as initiators of the chain oxidation of lipids. It is therefore interesting to eliminate these radicals from the headspace and the bulk of the food, as an alternative route to eliminating oxygen from the package. The radical scavenging efficiency of an antiradical substance depends on the rate of hydrogen atom abstraction from the phenyl group and also on the stability of the resulting radical. Strategies employed in active packaging generally include (1) the design of active compound releasing systems and (2) undesired compound scavenging systems [68]. In the first strategy, the generation of low molecular weight substances inside the packaging film and promoting their migration into the foodstuff is interesting. In the second strategy, one of the advantages of radical scavengers is their efficiency upon contact without need for release of active compounds. This has been shown for hydroxyl radicals in the gas phase scavenged by essential oils supported on silanized glass wool and active packaging films containing essential oils [69]. The principle has been applied in the examples of use of catechins in poly(ethylene terephthalate) [70] and catechins [71] or flavonoids in ethyl vinyl alcohol in using an adapted ORAC assay [72]. Although different methods exist to measure the radical scavenging activity of plastic films, the antiradical activity of the PLA-lignin film extracts was tested here with the help of the DPPH test, because it is readily available, largely used in literature and has already been employed for lignins [15]. Figure 3 shows the loss of the free radical DPPH $^{\circ}$ with time for different PLA-PROTOBIND 1000 samples immersed directly in the methanol/DPPH $^{\circ}$ solution. The neat PLA sample did not show any activity. Furthermore, it can be observed that the oxidative capacity of the film increased with increasing quantity of PROTOBIND 1000, as shown by the lower plateau value of the residual DPPH $^{\circ}$. Only 1 wt% PROTOBIND 1000 yielded very small decrease of the DPPH $^{\circ}$ concentration, and the signal was very noisy in the

beginning, while the 10 wt% PROTOBIND 1000 film reached a plateau value of 42 % remaining DPPH°. The films themselves seemed to show an antiradical activity. Interestingly, the PLA-PROTOBIND 1000 film sample shown in Fig. 3 was extracted for 24 h in ethanol/water (95/5 v/v). The same film sample was then immersed in the methanol/DPPH° solution a second time and the decrease of the DPPH° absorbance measured. It had still the possibility of oxidizing the DPPH° radical. This shows that a sustained protective action for food due to slow release kinetics might be possible. It is worth noting however that with the experimental setup scavenging effects on the

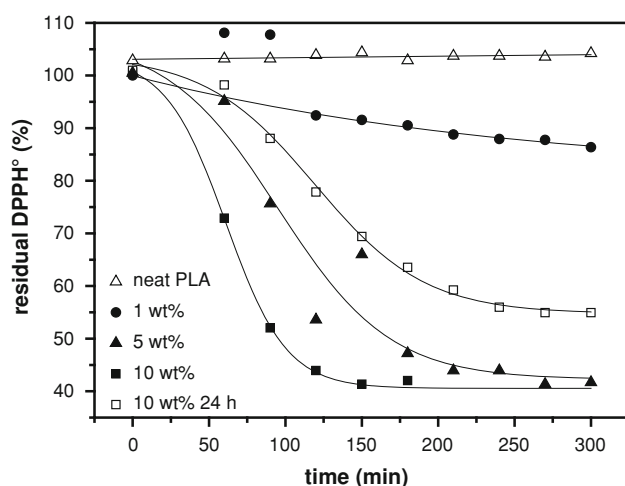


Fig. 3 Reaction kinetics of the free radical DPPH° during immersion of PLA-PROTOBIND 1000 films in the methanol solution. The sample indicated with 24 h was measured a second time after being extracted for 24 h in ethanol/water (95/5 v/v). Lines are drawn to guide the eye

surface cannot be clearly separated from effects due to surface release of phenolic compounds.

Therefore, the lignins and PLA-lignin films were immersed in a migration solution and the quantity of extracted phenolic compounds was assessed with the help of a semi-quantitative chromatography method (Fig. 4). In order to simulate fatty foodstuff, a solution of 95 % ethanol in water (v/v) was used as proposed by the European regulation for food contact materials [48]. The data in Fig. 4a confirmed that the content of possibly migrating free phenolic monomers in PROTOBIND 1000 is higher than in SARKANDA. The recovered quantity was much lower than the content of approximately 8 and 5 mg/g in PROTOBIND 1000 and SARKANDA, respectively, which was determined by Zheng et al. [8] with the help of a solvent extraction method. Ethanol/water is certainly a much less efficient extraction solvent compared to ethyl acetate/dichloromethane used by Zheng et al. [8]. The dashed line signifies the false detection level due to bleeding of the chromatography column. It can be observed in Fig. 4b that the migration from the PLA-lignin films with only 1 wt% of inclusion was not significant in the case of SARKANDA and very small in the case of PROTOBIND 1000. The detected quantities of free phenolic monomers in the migration solution formed two groups, where the higher group included only the PROTOBIND 1000 blends.

The antiradical activity of the different migrates from the PROTOBIND 1000 blends with increasing quantities of lignin was tested by the decrease of the concentration of the free radical of DPPH° in function of time (Fig. 5). The reaction kinetics of the PLA-1 wt% PROTOBIND 1000 film migrate was much slower than the kinetics of the 5 and 10 wt% PLA-PROTOBIND 1000 migrates because of the

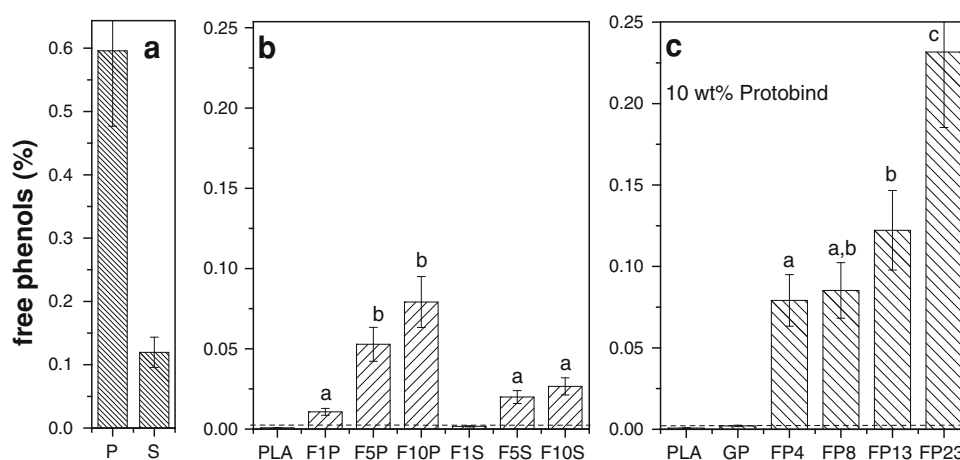


Fig. 4 Semi-quantitative gas chromatography determination of the migration of free phenolic monomers from PLA-lignin blends into ethanol/water (95/5 v/v) at room temperature for 24 h. The letters *a*, *b* and *c* correspond to statistical different groups. The dashed line marks the quantity detected due to column bleeding. Sample codes: In

b the codes indicate film samples with different quantities (*x* = 1, 5 or 10 wt%) of PROTOBIND 1000 (P) or SARKANDA (S). In **c** PLA-10 wt% PROTOBIND 1000 film samples are coded with the length of the heat treatment “FPt”, where *t* = 4, 8, 13 and 23 min. GP are the pellets of PLA-10 wt% PROTOBIND 1000

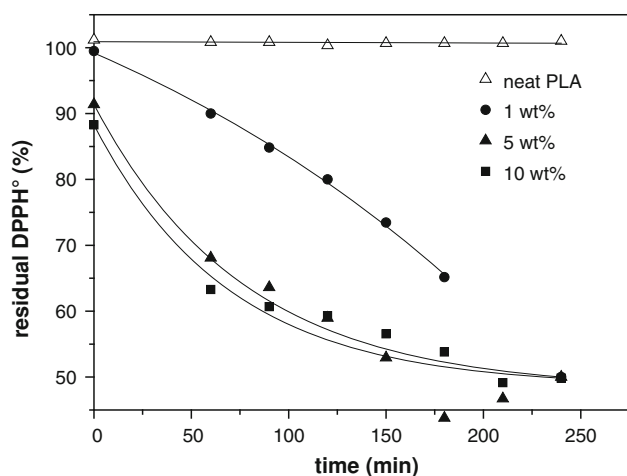


Fig. 5 Reaction kinetics of the free radical DPPH° with migrates of neat PLA (*open triangle*) and PLA-PROTOBIND 1000 samples in ethanol/water (95/5 v/v) for 24 h at room temperature. *Lines* are drawn to guide the eye

very small quantity of migrating substances, already observed in Fig. 4b. The inclusion of only 1 wt% lignin was therefore not enough to yield sufficient release from the matrix within the experimental time frame used to obtain an antiradical activity. No differences between the two higher concentrations were observed. The concentration of remaining DPPH° stabilized between around 50 % after 200 min. Kinetics are consistent with data given for slower reacting antioxidants like guaiacol [51]. The absolute plateau value of DPPH° cannot be compared with known substances, as it depends on the moles of antioxidant introduced. In the present case, the extract of the PLA-lignin films was a mixture of different substances and quantification of the free phenolic monomers was carried out only on the identified structures in the gas chromatogram. Therefore, the antiradical power of the migrate could not be determined.

In order to further investigate the effect of thermal processing on the release of free phenols, the film samples of PLA/PROTOBIND 1000 were thermocompressed for increasing times. The evolution in Fig. 4c showed that there is an augmentation in free phenolic compounds recovered in the migration solution which is correlated to the length of the thermal treatment. The samples separated into three groups where the sample treated 8 min corresponded to the middle group. The antiradical activity of the migration solutions of thermally treated PLA-lignin films was measured and results are plotted in Fig. 6. The plateau value of remaining DPPH° decreases with increasing thermal treatment. Two different groups were identified, the PLA-lignin pellets and the film treated for 4 min in one group (plateau values between 55 and 65 %) and the other films separated in a second group of higher activity

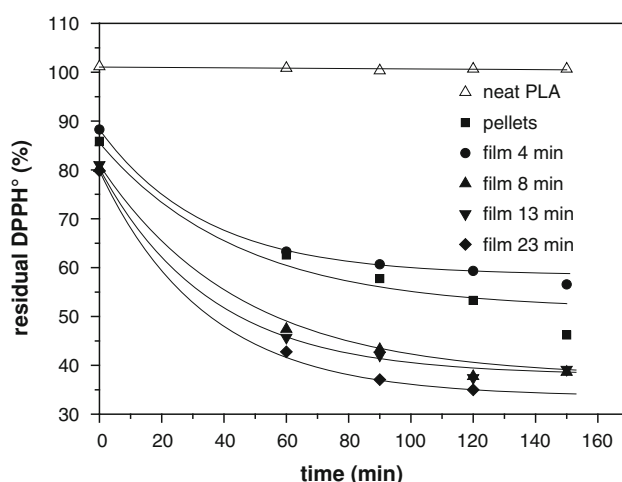


Fig. 6 Reaction kinetics of the free radical DPPH° with migrates in ethanol/water (95/5 v/v) of PLA-lignin samples containing 10 wt% lignin PROTOBIND 1000 after different heat treatments. *Lines* are drawn to guide the eye

(plateau values between 35 and 41 %). This result is consistent with the observation that the total quantity of the low molecular weight fraction of lignin increases with the thermal severity of processing (Fig. 4c). However, in Fig. 4c were identified 3 different groups, which were not found in the DPPH° test. Probably, the DPPH° test is more sensitive to variations in the quantity of antiradical substances. Furthermore, not all phenols have the same antioxidant power, which means that at in sum equal quantity differences in the antiradical efficiency can be expected. More detailed investigation of the quantity of each antiradical species will be required to deepen this question.

Conclusions

PLA-lignin films with two different sources of lignin, PROTOBIND 1000 and SARKANDA, were prepared by a two step twin screw extrusion and thermo-compression process and the characteristics of the composites were measured. Lignin and PLA were immiscible and distribution of lignin is homogeneous inside the matrix. Mechanical properties of the blends were only slightly diminished due to a counterbalancing effect between the reinforcement by the high molecular weight lignin and the plasticization by the low molecular weight lignin compounds and oxygen barrier properties were even slightly but significantly enhanced. The study of the lignin component revealed that the low molecular weight fraction of both lignins increased during processing. The immersion of the PLA-PROTOBIND 1000 film into the fatty food simulant yielded migration of radical scavengers into the solution as determined by the DPPH° test and the antiradical efficiency

increased with increasing severity of the heat treatment of the blends. This result opens the door to the fabrication of active packaging materials compounded with the natural antioxidant lignin as that they do not impact the mechanical and barrier performance of the polymer and even allow the creation of more active substances during the polymer processing. Further investigation of the kinetics of generation of antiradical substances and the identification of their respective antiradical power is now required to explore the feasibility of the process.

Acknowledgments The authors acknowledge the scientific research fund of AgroParisTech for funding.

References

- Boudet AM, Lapierre C, Grimapettenati J (1995) *New Phytol* 129:203
- Stewart D (2008) *Ind Crops Prod* 27:202
- Doherty WOS, Mousavioun P, Fellows CM (2011) *Ind Crops Prod* 33:259
- Lora JH, Glasser WG (2002) *J Polym Environ* 10:39
- Graupner N (2008) *J Mater Sci* 43:5222
- Baumberger S, Lapierre C, Monties B (1998) *J Agric Food Chem* 46:2234
- Baumberger S, Lapierre C, Monties B, Della Valle G (1998) *Polym Degrad Stab* 59:273
- Zheng D, Méchin V, Pollet B, Dole P, Coqueret X, Baumberger S (2008) In: 10th EWLP, European Workshop on Lignocellulosics and Pulp, KTH, Royal Institute of Technology, Stockholm, pp 380–383
- Reti C, Casetta M, Duquesne S, Bourbigot S, Delobel R (2008) *Polym Adv Technol* 19:628
- Toh K, Nakano S, Yokoyama H, Ebe K, Gotoh K, Noda H (2005) *Polym J* 37:633
- Pucciariello R, D'Auria M, Villani V, Giammarino G, Gorrasi G, Shulga G (2010) *J Polym Environ* 18:326
- Cruz JM, Dominguez JM, Dominguez H, Parajo JC (2001) *J Agric Food Chem* 49:2459
- Ugartondo V, Mitjans M, Vinardell MP (2008) *Bioresour Technol* 99:6683
- Vinardell MP, Ugartondo V, Mitjans M (2008) *Ind Crops Prod* 27:220
- Dizhbite T (2004) *Bioresour Technol* 95:309
- Pan XJ, Kadla JF, Ehara K, Gilkes N, Saddler JN (2006) *J Agric Food Chem* 54:5806
- Lu Q, Liu WJ, Yang L, Zu YG, Zu BS, Zhu MH, Zhang Y, Zhang XN, Zhang RR, Sun Z, Huang JM, Zhang XN, Li WG (2012) *Food Chem* 131:313
- Faustino H, Gil N, Baptista C, Duarte AP (2010) *Molecules* 15:9308
- Garcia A, Toledano A, Andres MA, Labidi J (2010) *Process Biochem* 45:935
- Karvela E, Makris DP, Kefalas P, Moutounet M (2008) *Food Chem* 110:263
- Li M-F, Sun S-N, Xu F, Sun R-C (2012) *J Agric Food Chem* 60:1703
- Alexy P, Kosikova B, Crkonova G, Gregorova A, Martis P (2004) *J Appl Polym Sci* 94:1855
- Gregorova A, Kosikova B, Stasko A (2007) *J Appl Polym Sci* 106:1626
- Mengelloglu F, Karakus K (2008) *Fresenius Environ Bull* 17:211
- Samal SK, Fernandes EG, Corti A, Chiellini E (2009) *Int J Mater Prod Technol* 36:62
- Pouteau C, Baumberger S, Cathala B, Dole P (2004) *C R Biol* 327:935
- Dinis TCP, Madeira VMC, Almeida LM (1994) *Arch Biochem Biophys* 315:161
- Frankel EN (1996) *Food Chem* 57:51
- Shahidi F (2000) *Nahrung Food* 44:158
- Bentayeb K, Rubio C, Sanchez C, Battle R, Nerin C (2007) *Ital J Food Sci* 110
- Ugartondo V, Mitjans M, Pilar Vinardell M (2009) *Ind Crops Prod* 30:184
- Berlinet C, Ducruet V, Brillouet JM, Reynes M, Brat P (2005) *Food Addit Contam Part A Chem Anal Control Expo Risk Assess* 22:185
- Saint-Eve A, Levy C, Le Moigne M, Ducruet V, Souchon I (2008) *Food Chem* 110:285
- Lopez-de-Dicastillo C, Gomez-Estaca J, Catala R, Gavara R, Hernandez-Munoz P (2012) *Food Chem* 131:1376
- Pereira de Abreu DA, Maroto J, Rodriguez KV, Cruz JM (2012) *J Sci Food Agric* 92:427
- Nerin C, Tovar L, Djenane D, Camo J, Salafranca J, Beltran JA, Roncales P (2006) *J Agric Food Chem* 54:7840
- Wessling C, Nielsen T, Giacini JR (2001) *J Sci Food Agric* 81:194
- Van Bree I, De Meulenaer B, Samapundo S, Vermeulen A, Ragaert P, Maes KC, De Baets B, Devlieghere F (2010) *Innov Food Sci Emerg Technol* 11:511
- Hambleton A, Voilley A, Debeaufort F (2012) *Food Hydrocoll* 25:1128
- Karbowiak T, Debeaufort F, Voilley A, Trystram G (2009) *Innov Food Sci Emerg Technol* 10:116
- Antunez PD, Omary MB, Rosentrater KA, Pascall M, Winstone L (2012) *J Food Sci* 77:S1
- Cardona ED, Noriega MD, Sierra JD (2012) *J Plast Film Sheet* 28:63
- Soto-Valdez H, Auras R, Peralta E (2011) *J Appl Polym Sci* 121:970
- Manzanarez-Lopez F, Soto-Valdez H, Auras R, Peralta E (2011) *J Food Eng* 104:508
- Hwang SW, Shim JK, Selke SE, Soto-Valdez H, Matuana L, Rubino M, Auras R (2012) *Polym Int* 61:418
- Ouyang WZ, Huang Y, Luo HJ, Wang DS (2012) *J Polym Environ* 20:1
- Li JC, He Y, Inoue Y (2003) *Polym Int* 52:949
- 97/48/EC D, Directive 97/48/EC of the European Commission. Official Journal of the European Communities (29 July 1997)
- Fischer E, Sterzel H, Wegner G (1973) *Kolloid-Zeitschrift Zeitschrift für Polymere* 251:980
- Gellerstedt G (1992) In: Lin Y, Dence C (eds) *Methods in lignin chemistry*. Springer-Verlag, Berlin, pp 485–523
- Brand Williams W, Cuvelier ME, Berset C (1995) *Food Sci Technol Lebensm Wiss Technol* 28:25
- Lim LT, Auras R, Rubino M (2008) *Prog Polym Sci* 33:820
- Salazar R, Domenek S, Courgneau C, Ducruet V (2012) *Polym Degrad Stab* 97:1871
- Courgneau C, Domenek S, Lebosse R, Guinault A, Averous L, Ducruet V (2012) *Polym Int* 61:180
- Pillin I, Montrelay N, Grohens Y (2006) *Polymer* 47:4676
- Domenek S, Courgneau C, Ducruet V (2011) In: Kalia S, Averous L (eds) *Biopolymers: biomedical and environmental applications*. John Wiley & Scrivener Pub., pp 183–223
- Martino VP, Ruseckaite RA, Jiménez A (2009) *Polym Int* 58:437
- Elangovan D, Yuzay IE, Emselke S, Auras R (2011) *Polym Int* 61:30
- Auras R, Harte B, Selke S (2004) *Macromol Biosci* 4:835

-
60. Colomines G, Ducruet V, Courgneau C, Guinault A, Domenek S (2010) *Polym Int* 59:818
 61. Auras RA, Harte B, Selke S, Hernandez R (2003) *J Plast Film Sheet* 19:123
 62. Guinault A, Sollogoub C, Ducruet V, Domenek S (2012) *Eur Polym J* 48:779
 63. Drieskens M, Peeters R, Mullens J, Franco D, Lemstra PJ, Hristova-Bogaerds DG (2009) *J Polym Sci Part B Polym Phys* 47:2247
 64. Sawada H, Takahashi Y, Miyata S, Kanehashi S, Sato S, Nagai K (2010) *Trans Mater Res Soc Jpn* 35:241
 65. Courgneau C, Domenek S, Guinault A, Averous L, Ducruet V (2011) *J Polym Environ* 19:362
 66. Baumberger S, Abaecherli A, Fasching M, Gellerstedt G, Goselink R, Hortling B, Li J, Saake B, de Jong E (2007) *Holzfor-schung* 61:459
 67. López-de-Dicastillo C, Pezo D, Nerín C, López-Carballo G, Catalá R, Gavara R, Hernández-Muñoz P (2012) *Packag Technol Sci*. doi:[10.1002/pts.992](https://doi.org/10.1002/pts.992)
 68. Nerin C, Tovar L, Salafranca J (2008) *J Food Eng* 84:313
 69. Pezo D, Salafranca J, Nerin C (2008) *J Chromatogr A* 1178:126
 70. Colon M, Nerin C (2012) *J Agric Food Chem* 60:9842
 71. Lopez de Dicastillo C, Nerin C, Alfaro P, Catala R, Gavara R, Hernandez-Munoz P (2011) *J Agric Food Chem* 59:7832
 72. Bentayeb K, Vera P, Rubio C, Nerin C (2009) *Anal Bioanal Chem* 394:903