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Stéphane URCUN, Giuseppe SCIUMÈ, Stéphane P.A. BORDAS, Pierre-Yves ROHAN - Cortex tissue relaxation and slow to medium load rates dependency can be captured by a two-phase flow poroelastic model - Journal of the Mechanical Behavior of Biomedical Materials - Vol. 126, p.104952 - 2021

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# Cortex tissue relaxation and slow to medium load rates dependency can be captured by a two-phase flow poroelastic model

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## Abstract

This paper investigates the complex time-dependent behavior of cortex tissue, under adiabatic condition, using a two-phase flow poroelastic model. Motivated by experiments and Biot's consolidation theory, we tackle time-dependent uniaxial loading, confined and unconfined, with various geometries and loading rates from  $1 \mu\text{m/s}$  to  $100 \mu\text{m/s}$ . The cortex tissue is modeled as the porous solid saturated by two immiscible fluids, with dynamic viscosities separated by four orders, resulting in two different characteristic times. These are respectively associated to interstitial fluid and glial cells. The partial differential equations system is discretised in space by the finite element method and in time by Euler-implicit scheme. The solution is computed using a monolithic scheme within the open-source computational framework FEniCS. The parameters calibration is based on Sobol sensitivity analysis, which divides them into two groups: the tissue specific group, whose parameters represent general properties, and sample specific group, whose parameters have greater variations. Our results show that the experimental curves can be reproduced without the need to resort to viscous solid effects, by adding an additional fluid phase. Through this process, we aim to present multiphase poromechanics as a promising way to a unified brain tissue modeling framework in a variety of settings.

*Keywords:* poromechanics, brain tissue, mechanical testing

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## 1. Introduction

The biomechanical characterization of human brain tissue and the development of appropriate mechanical models is crucial to provide realistic computational predictions. These predictions can assist in understanding the mechanical environment involved in neurodevelopment and neurological disorders [1, 2], in simulating traumatic brain injury, to investigate the mechanical pathogenesis of head trauma [3] and in studying head injuries and developing protection systems [4]. Mathematical modeling is also the key to devising brain surgery simulation for training, assistance and guidance [5, 6].

From an experimental perspective, several sophisticated mechanical tests have been proposed and conducted both on human and animal brain tissue in the past decades (see [7] for a complete review). These have consistently shown that brain tissue is non-linear, asymmetric in tension-compression and sensitive to loading rates. Moreover, grey and white matter, when they are isolated, exhibit different stiffness and brain tissue can be separated into four regions, with their own mechanical profile: cortex, corona radiata, basal ganglia and corpus callosum. It has been shown that the mechanical response of brain tissue is sensitive to the time scale and the characteristic length of loading. This has been attributed by several authors to the ultra-soft, gel-like nature of brain tissue, which implies that effects on very

small length and time scales may hold an important impact on the overall mechanical behavior [7].

To capture the mechanical response of brain tissue, several constitutive models have been proposed based on the type and range of the strain rates associated with pathological and normal conditions. In particular, extensive research has been conducted for brain matter experiencing compression at quasi-static loading [8, 9]. These studies have shaped our understanding of the hyperelastic time-independent response of human brain tissue. However, its time-dependent behavior at finite strains and under various loading conditions remains insufficiently understood.

Visco-elastic models [10, 11, 12, 13] are typically chosen to reproduce the time-dependent, hysteresis, preconditioning softening (regarding this topic, see the review [7] section 3.5) as well as the stress relaxation observed in experiments. The coupling of large deformation (usually hyper-elastic) and visco-elasticity has allowed to accurately capture various deformation types in a large range of strain. These have been employed to predict the essential features of brain tissue: non-linearity, hysteresis, and tension-compression asymmetry. Yet, most models developed to date have been tailored to reproduce particular loading scenarios or for specific applications. We conclude that this specific kind of models, that could be denoted as phenomenological, do not contain the required components to be transferable from one type of experimental conditions to another. Our aim is to alleviate these deficiencies, so that models

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developed for one patient can be used for another patient, and to make this possible for the most varied types of boundary and environmental conditions. We propose in this paper a particular multi-phase poro-elastic model. We show that this model, once calibrated on a certain set of experiments, can also reproduce the behaviour of brain tissue observed in completely different loading conditions. We therefore hypothesise that such a model could be a sound starting point for a generic mechanical model of the brain. In the appendix, the interested reader may also find the indication that, for a viscoelastic model, to mimic the behavior of a porous material, the parameters must be adjusted according to the specimen size.

Several studies [14, 15, 16], based on a series of consolidation tests, suggested that extracellular fluid flow dominates the apparent viscoelastic properties of brain tissue. Based on mixture theory, poroelasticity has been extensively used to model brain tissue [17, 18, 19, 20, 21]. Poroelastic media mechanics, [22], which describe the mechanical behavior of a porous solid containing viscous fluid, are compatible with processes which involve low-strain-rate deformations of the brain, such as edema, hydrocephalus, hemorrhage [23, 24] and infusion [25, 26]. However, single phase flow poromechanics show limitation to render the highly non-linear behavior of the brain tissue.

Franceschini *et al.* (2006) [14] conclude that consolidation (deformation of the solid matrix due to pore fluid flow and its drainage from the interstitial space) is the leading mechanism in quasi-static deformation of brain tissue. To achieve complete adherence between their experimental data and their theoretical formulations, they conclude that the addition of a viscous component to the rheological model is necessary.

This last point remains a matter of debate. Budday *et al.* [7] hypothesised that brain tissue rheology is characterized by at least two different timescales, which are attributed to viscous and porous effects, respectively. The viscous response can be related to the intracellular interactions within the network of cells forming the solid phase of the tissue, while the porous medium behavior is associated with the interaction between the solid and fluid phase. In a recent study, Comellas *et al.* [27] explored in details the interplay between single phase poro-elasticity and finite visco-elasticity. Their model is capable of accurately reproducing consolidation, shear stress and cycle loading tests. However, the respective influence of porous and viscous components are strongly dependent on the parameters calibration, which could impede the physical interpretation of the results.

Unlike conventional approaches that combine viscoelastic and poroelastic behaviors into a single framework [28], we hypothesize in this article that a two-phase flow poroelasticity can also capture the stress relaxation and the sensitivity of a set of selected load rates. The two fluid phases, with their respective dynamic viscosities, allow for reproducing the two different time scales. As a first step, we limited our study on the cortex region of the brain, which is known to be soft and isotropic [7]. We first reproduce the experimental results of Franceschini *et al.* [14] without the use of a viscoelastic solid and, with the same model, we reproduce the experimental set of Budday *et al.* [29] composed of various indenters, load rates and stress

relaxation.

## 2. Experimental data

### Consolidation tests

The consolidation test (an uniaxial strain under free drainage conditions) is crucial for poromechanical modeling, as this strain can not be reproduced by an elastic law and is not adapted to viscoelastic models [14]. In this contribution we chose to reproduce the experiment of Franceschini conducted on  $N=12$  specimens. The experimental procedure is briefly reported here. For more details the interested reader could refer to [14].

Twelve human parietal lobe cylindrical samples of 30 mm diameter and 5 to 8 mm height are harvested, within 12h after death, and placed into a consolidometer specifically designed for the tests, shown in Fig. 1A. The top and bottom of the samples are under free drainage conditions, using a filter paper against a porous brass employed in geotechnical consolidometer.

We identify three different profiles on the twelve samples, where the same initial load (6 Newton) will lead deformation from 3.5% to 6.5%. We retrieve the same three profiles on the 3 Newton load series. As a consequence, we select six samples from [14] for this study, one of each profile for the 3 and 6 Newton one step loads. We adopt the following nomenclature for the samples:  $XN$  for  $X$  Newton load series, followed by the figure index and letter of [14], *e.g.* 3N C2 a.

### Indentation tests

Due to the difficulties in producing consistent samples, and due to the alterations to the micro-structural arrangement during the sample preparation, it has been reported that stiffness values using confined compression creep and stress relaxation tests are often not reproducible and may vary by an order of magnitude or more. Indentation has been shown to provide a robust, reliable, and repeatable method to quantify the mechanical properties *in situ* without altering the local micro-architectural arrangement.

Four indentation tests on *ex vivo* bovine grey matter, within 6 hours after death, are reproduced. These are subsets of the long range indentation type, on  $N = 192$  samples, described in [29]. Before any test, all samples are subjected to a  $100\ \mu\text{m}$  indentation depth to stabilize the material response and ensure parallelism. The experimental design is shown in Fig. 2 A and the characteristics of the tests can be found in Table 1. Budday *et al.* present indentation tests with loading rates between  $1\ \mu\text{m/s}$  and  $100\ \mu\text{m/s}$ , for reason of brevity, we choose to reproduce these extreme values without the intermediate  $10\ \mu\text{m/s}$  and  $20\ \mu\text{m/s}$ . In [29], Budday *et al.* present mean experimental curves for each test and indicate 21% of standard deviation in the set of samples.

## 3. Mathematical Modelling

The descriptions of cortex tissue as a porous medium can take several forms: a solid scaffold saturated by a fluid (Hakim

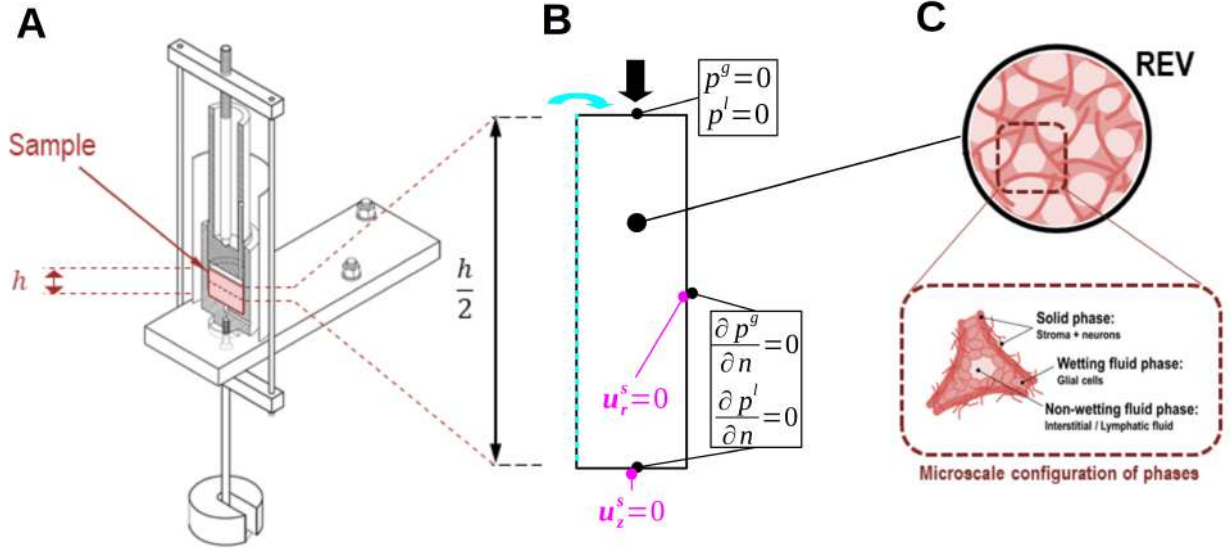


Figure 1: **Consolidation tests on *ex vivo* human cortex [14].** **A** Experimental design of consolidometer of Franceschini *et al.* [14] (reproduced with the authorisation of the authors). **B** Boundary conditions of the physical problem: the fluids are free to escape to loaded boundary (drained condition), the other boundaries are impervious; except at the loaded boundary, the displacement of the solid scaffold is allowed only on the tangential direction (slip condition). **C** Representative elementary volume (REV) with the phase components of the model framework.

Table 1: **Characteristics of the indentation tests on *ex vivo* bovine cortex [29].** Test 1, 2 and 3 belong to long range indentation tests, Test 4 is a relaxation test.

Characteristics	Test 1	Test 2	Test 3	Test 4
Indentation depth $\mu\text{m}$	300	300	300	100
Load rate $\mu\text{m/s}$	5	5	1	100
Indenter $\varnothing$ mm	1	0.75	0.75	1.5
Unloading of 300 $\mu\text{m}$	Yes	Yes	Yes	No
Holding of 600 s	No	No	No	Yes

*et al.* 1976), a visco-elastic scaffold saturated by a fluid [14], an elastic scaffold separated into two compartments, vascular and extra-vascular, each saturated by a different fluid [30], among others. In this study, we focus on *ex vivo* material, so the vascular activity will be neglected and the vascular network will be considered a part of the solid scaffold. Within the parenchyma, we distinguish two fluid phases: glial  $g$  phase, for which we prescribed a dynamic viscosity close to gel-like, and interstitial/lymphatic  $l$  phase, with a prescribed dynamic viscosity one order higher than water. A summary of the representative elementary volume is shown in Fig. 1C. This specific description of cortex tissue by two immiscible fluids within a porous solid has no experimental equivalent. Therefore, we present the quantitative information to set the initial guess of the parameters from several sources in the experimental literature.

### Governing equations

A porous medium consists in a solid scaffold, that is to say all the material that act as structural solids in the porous medium. Its volume fraction is denoted  $\varepsilon^s$ . The porosity is the volume

fraction saturated by the fluids, denoted  $\varepsilon$ , and

$$\varepsilon^s + \varepsilon = 1 \quad (1)$$

We distinguish, in this contribution, the glial phase  $g$  from the remaining fluid phase  $l$ . These two phases are assumed to be immiscible fluids with their respective saturation  $S^l$  and  $S^g$  and

$$S^l + S^g = 1 \quad (2)$$

The volume occupied by the liquid phase and by the glial phase are respectively denoted  $\varepsilon S^l$  and  $\varepsilon S^g$ .

The mass balance of the porous solid reads:

$$\frac{D^s}{Dt} (\rho^s \varepsilon^s) + \rho^s \varepsilon^s \nabla \cdot \mathbf{v}^s = 0 \quad (3)$$

- with  $\frac{D^s}{Dt} (\rho^s \varepsilon^s)$  the evolution of the solid scaffold mass
- and  $\rho^s \varepsilon^s \nabla \cdot \mathbf{v}^s$  its flux term.

The mass balance of the fluids phases reads:

$$\frac{D^s}{Dt} (\rho^g \varepsilon S^g) + \nabla \cdot (\rho^g \varepsilon S^g \mathbf{v}^{\bar{g}s}) + \rho^g \varepsilon S^g \nabla \cdot \mathbf{v}^s = 0 \quad (4)$$

$$\frac{D^s}{Dt} (\rho^l \varepsilon S^l) + \nabla \cdot (\rho^l \varepsilon S^l \mathbf{v}^{\bar{l}s}) + \rho^l \varepsilon S^l \nabla \cdot \mathbf{v}^s = 0 \quad (5)$$

- with  $\frac{D^s}{Dt} (\rho^\alpha \varepsilon S^\alpha)$  the evolution of the fluid phase  $\alpha$
- $\nabla \cdot (\rho^\alpha \varepsilon S^\alpha \mathbf{v}^{\bar{\alpha}s})$  the flux term of the phase  $\alpha$  where  $\mathbf{v}^{\bar{\alpha}s}$  is the phase  $\alpha$  velocity relatively to the solid:  $\mathbf{v}^{\bar{\alpha}s} = \mathbf{v}^{\bar{\alpha}} - \mathbf{v}^s$
- and  $\rho^\alpha \varepsilon S^\alpha \nabla \cdot \mathbf{v}^s$  the contribution of the solid deformation to the mass conservation of the phase  $\alpha = l, g$

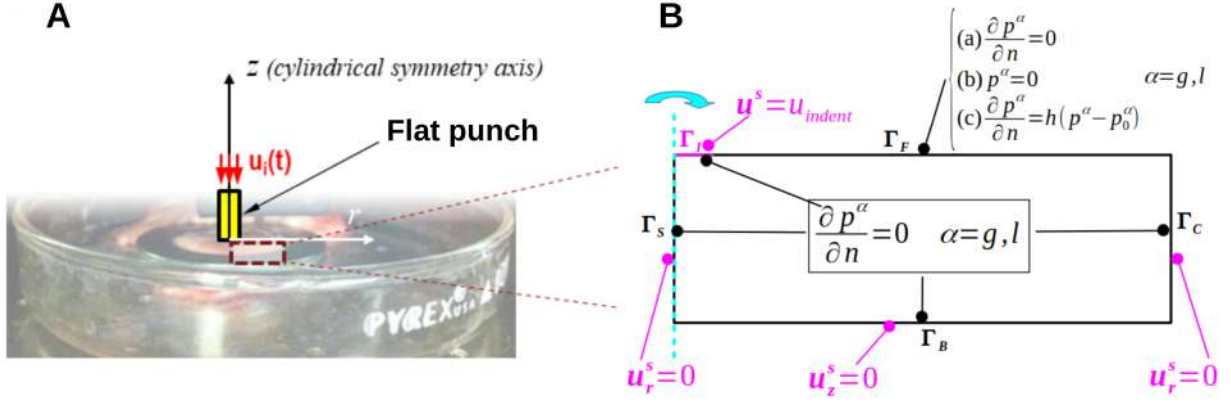


Figure 2: **Indentation test on *ex vivo* bovine cortex** [29]. **A** Experimental design of indentation tests of Budday *et al.* [29] (reproduced with the authorisation of the authors). The sample is considered under cylindrical symmetry hypothesis,  $\mathbf{u}_i(t)$  is displacement prescribed by the indenter on the sample surface.

**B** Boundary conditions of the physical problem in cylindrical axis symmetry. Boundaries are denoted  $\Gamma_S$  the axis of symmetry,  $\Gamma_B$  and  $\Gamma_C$  the slip boundaries prescribed in the experimental design,  $\Gamma_I$  the indented surface and  $\Gamma_F$  the free surface, where three different boundary conditions are tested.

### Constitutive relationships

The glial phase will be denoted as the wetting phase, that is to say, the phase that preferentially covers the solid scaffold. Therefore, it is the one that sustains the mechanical load before transmitting it to the non-wetting phase, namely the liquid phase. The pressure relationship with the phases saturation is justified by the simplified entropy inequality of the thermodynamically constrained averaging theory (TCAT) [31]. At the microscale equilibrium, it is assumed that the evolution of the interface surface is negligible compared to the evolution of the fluid saturation, therefore the interfacial tension between the phases is directly linked to the pressure difference (see [32]). This implies a pressure jump between the wetting phase  $g$  and the non-wetting phase  $l$ , whose saturation  $S^l$  depends on the pressure jump  $p^{lg} = p^l - p^g$  (see Eq.6).

From [32], the expression of the liquid phase saturation, dependent on the difference of the phases pressure, is adapted from the heuristic formulation of van Genuchten [33]:

$$S^l = \frac{2}{\pi} \arctan\left(\frac{p^{lg}}{a}\right) \quad (6)$$

with the theoretical parameter  $a$ , which represents the intensity of the interplay between the solid scaffold and the non-wetting phase. Its range commonly taken is [500; 800] (see [34], [35], [36], [37]).

The solid scaffold has its intrinsic permeability  $k_{\text{int}}^s$  and follows linear elasticity (Young modulus  $E$ , Poisson's ratio  $\nu$ ):

$$\bar{\sigma}_{\text{eff}} = \lambda \text{tr}(\bar{\epsilon}) \mathbb{I} + 2\mu \bar{\epsilon} \quad (7)$$

with  $\mathbb{I}$  the identity tensor,  $\bar{\epsilon}(\mathbf{u}^s) = \frac{1}{2}(\nabla \mathbf{u}^s + (\nabla \mathbf{u}^s)^T)$  the linearised strain tensor, and the Lamé constants  $\lambda = \frac{E\nu}{(1+\nu)(1-2\nu)}$  and  $\mu = \frac{E}{2(1+\nu)}$ .

The two fluid phases have their own relative permeabilities:

$$k_{\text{rel}}^l = (S^l)^{A_l} \quad k_{\text{rel}}^g = (S^g)^{B_g} \quad (8)$$

with  $A_l$  and  $B_g$  to be calibrated.

The interaction between fluid phases and the solid scaffold are modeled by a generalized Darcy's flow, deduced from the linear momentum conservation of fluid phases:

$$-\frac{k_{\text{rel}}^{\alpha} k_{\text{int}}^s}{\mu^{\alpha}} \nabla p^{\alpha} = \varepsilon S^{\alpha} (\mathbf{v}^{\alpha s}) \quad \alpha = g, l \quad (9)$$

where  $k_{\text{int}}^s$  is the intrinsic permeability of the solid scaffold,  $\mu^{\alpha}$ ,  $k_{\text{rel}}^{\alpha}$  and  $p^{\alpha}$  are respectively the dynamic viscosity, relative permeability and the pressure of each fluid phase  $\alpha = l, g$ .

### Final system of governing equations

The physical system is governed by the mass conservation of each phase and by linear momentum conservation of the overall system. The primary unknowns of the model are the solid scaffold displacement  $\mathbf{u}^s$ , the glial phase pressure  $p^g$  and the liquid phase pressure difference  $p^{lg}$ .

Eqs.3-5 are treated as follows:

- Eq.3 is injected into Eqs.4-5 thanks to the constraint Eq.1
- the difference of order between the mechanical constrains and the compressibility of the components allows us to neglect the variation of their density, therefore  $\rho^{\alpha}$  is considered constant  $\alpha = s, g, l$ . We also assume that the component have the same density  $\rho^s = \rho^g = \rho^l$
- the evolution of the glial volume fraction  $\varepsilon \frac{D^s S^l}{Dt}$  is expressed with respect to the constraint Eq.2. Its evolution depends on the interstitial fluid pressure  $S^l(p^{lg})$ , see the constitutive relationship Eq.6

We obtain the following system:

$$\begin{cases} -\varepsilon \frac{dS^l}{dp^{lg}} \frac{D^s p^{lg}}{Dt} - \nabla \cdot \left[ \frac{k_{rel}^s k_{int}^s}{\mu^s} \nabla p^g \right] + S^g \nabla \cdot \mathbf{v}^s = 0 \\ \varepsilon \frac{dS^l}{dp^{lg}} \frac{D^s p^{lg}}{Dt} - \nabla \cdot \left[ \frac{k_{rel}^l k_{int}^s}{\mu^l} \nabla (p^g + p^{lg}) \right] + S^l \nabla \cdot \mathbf{v}^s = 0 \end{cases} \quad (10)$$

The governing equations are completed with the momentum balance of the system:

$$\nabla \cdot \sigma_T = \mathbf{0} \quad \text{with} \quad \sigma_T = \sigma_{\text{eff}} - \beta p^s \quad (11)$$

where  $\sigma_{\text{eff}}$ , the effective stress described Eq.7, represents the solid contribution to the mechanical stress.  $\beta$  is the Biot's coefficient, set to 1 thanks to the hypothesis of the phases incompressibility.  $p^s = S^l p^l + S^g p^g$ , denoted the solid pressure, represents the fluids' contribution to the mechanical stress.

*Assumption on the porous medium components and parameters motivated by literature*

The stiffness of the cortex has been intensively studied ([38, 39, 40], among many others). It both depends on the chosen modeling framework and the sample composition, so the values of the Young's modulus  $E$  can widely vary. In time-independent modeling, as linear elastic or hyperelastic, using linear elastic law leads to scatter values Young's modulus  $E$ , from 2.5 kPa [41] to 8 kPa [38]. Non-linear hyperelastic modeling has more homogeneous values, with  $E$  between 1.1 kPa [39] and 3 kPa [40]. In the reproduced experimental study [29], the authors found by contact theory  $E = 1.389 \pm 0.289$  kPa. Conversely, the values of Poisson's ratio  $\nu$  are quite consensual in the literature, as the material is constantly described as nearly incompressible, with values varying between 0.495 [38] and 0.45 [42].

In Barnes *et al.* [43], the extra-cellular matrix (ECM) volume fraction is estimated to be 20%. In Lei *et al.* [44], the volume fraction of the interstitial fluid (IF), added to the ECM, is estimated to be between 15% and 20%. From Bender and Klose [45], the IF fraction, strictly limited to the parenchyma, is estimated to be between 5% and 10%, and the vascular system (in [44]) between 3% and 5%. Finally, the distribution indicated by Azevedo *et al.* in [46] for the cerebral cortex gives 20% neuron, 80% non-neuron with  $\pm 2\%$  of uncertainty.

We define the solid scaffold as the sum of the stroma cells (membrane, epithelial and endothelial cells), the ECM, the vascular system - because blood circulation is not considered in *ex vivo* testing - and the neurons (bodies and axons). Based on the estimation provided above, we obtain the following range of values for the solid scaffold fraction: the minimum at  $(0.15 - 0.1) + 0.03 + 0.18 = 0.24$  and the maximum at  $0.2 + 0.1 + 0.05 + 0.22 = 0.57$ . Its complement, the porosity  $\varepsilon$ , is within the range  $0.43 \leq \varepsilon \leq 0.76$ .

As written in [45], the IF volume fraction estimated to  $0.05 \leq \varepsilon S^l \leq 0.1$ . With the estimated range of porosity  $\varepsilon$ , we obtain for the liquid phase saturation:  $0.065 \leq S^l \leq 0.232$ ; and its complement, the glial phase saturation:  $0.828 \leq S^g \leq 0.935$ .

As initial range of values for the liquid phase dynamic viscosity  $\mu_l$ , we keep different estimation provided in [47] for cerebral fluid, between  $0.7 \cdot 10^{-3}$  Pa · s and  $1 \cdot 10^{-3}$  Pa · s. For the rheology of the glial phase, as we do not have experimental data, we choose a value close to literature used for generic cells (see [34] and [37])  $\mu_g \approx 30$  Pa · s.

Regarding the intrinsic permeability  $k_{int}^s$  of the solid scaffold, there is still a large knowledge gap. The important difficulty to design experiment on living tissue and the preferential use of single phase solid mechanics rather than poromechanics are probably partly responsible for this problem. In 2021, Sowinski *et al.* [47] reported ranges of values for hydraulic conductivity  $K$ , using *in silico* magnetic resonance elastography. They reported hydraulic conductivity values from  $2 \cdot 10^{-11}$  ms<sup>-1</sup> to  $2 \cdot 10^{-10}$  ms<sup>-1</sup>. Same year, Jamal *et al.* in [48], reported range of values for the intrinsic permeability  $k_{int}^s$ , by *ex vivo* perfusion experiment. They reported a  $k_{int}^s$  mean value of  $10^{-16}$  m<sup>2</sup>, with a strong influence of tissue anisotropy, from  $2 \cdot 10^{-17}$  m<sup>2</sup> to  $3.2 \cdot 10^{-15}$  m<sup>2</sup>, with one order of magnitude difference if perfusion is parallel or perpendicular to the white matter fibers. Retrieve intrinsic permeability  $k_{int}^s$  from hydraulic conductivity  $K$  is not straightforward. If the ratio  $\frac{k_{int}^s}{\mu}$  is common to the different equivalence, the size of the sample, its density and difference of pressure could be at play. Applying the ratio on hydraulic conductivity, a dynamic viscosity  $\mu$  with the order of  $1 \cdot 10^{-3}$  Pa · s will lead to an intrinsic permeability  $k_{int}^s$  between  $2 \cdot 10^{-14}$  m<sup>2</sup> and  $2 \cdot 10^{-13}$  m<sup>2</sup>, which is two orders higher than [48]. In [47], Sowinski *et al.* hypothesise that *in vitro* experiments tend to lead to smaller values, due to pore collapse in sampled tissue. In [48], Jamal *et al.* claim that these microstructural changes become significant after 6h post-mortem, hence they perform their experiments within these 6h. As we do not have sufficient information, we have no other choice to keep this large range, from  $10^{-17}$  to  $10^{-13}$ , as acceptable values of  $k_{int}^s$ .

#### 4. Computational framework

Using the FEniCS python libraries, the solution of the problem has been done with the finite element method, with a monolithic solution process. Boundary conditions for the consolidation tests are shown in Fig.1B and for the indentation tests in Fig.2B. The computations have been run on a mesh of  $4 \times 20$  elements for consolidation tests and  $98 \times 60$  for indentation tests, both are cylindrical axis symmetric representation of the sample. The same computations have been run with meshes with three levels of refinement, with the same results.

The boundaries of the consolidation tests are shown in Fig.1B. The fluids are free to escape to loaded boundary (drained condition) and the other boundaries are impervious. For the solid scaffold, except at the loaded boundary, its displacement is allowed only on the tangential direction (slip condition). The boundaries of the indentation tests are shown in Fig.2B. Its boundary conditions are denoted by  $\Gamma_S$  the axis of symmetry,

by  $\Gamma_B$  and  $\Gamma_C$  the slip boundaries prescribed in the experimental design.  $\Gamma_I$  is the indented surface, where the fluid is assumed under impervious condition and  $\Gamma_F$  the free surface.

All the codes used for computation are available on Github, at <https://github.com/StephaneUrcun/BrainTissue>

#### Local sensitivity analysis and parameters calibration

As an initial guess, we first set all the parameters within their ranges given in section 3 paragraph 'Assumption motivated by literature'. Seven are denoted *tissue specific*, because they are general properties, more likely to be shared by all samples: the porosity  $\varepsilon$ , the Poisson's ratio  $\nu$ , the dynamic viscosity of the two fluid phases  $\mu_l, \mu_g$ , their corresponding tortuosity exponent  $A_l, B_g$  and  $a$ , the referenced pressure of the cell-ECM interplay. Young's Modulus  $E$  and intrinsic permeability  $k_{\text{int}}^s$ , which have the wider ranges, and the initial liquid phase saturation  $S_0^l$  are denoted *sample specific* ( $S_0^l$  varies for each sample, but within the prescribed physiological range, see section 3 paragraph 'Assumption motivated by literature').

We perform a variance-based local sensitivity analysis on the parameters at their initial guess value  $\pm 10\%$ , described Appendix B. We perform first order sensitivity analysis (the parameters are modified one at a time) as well as second order analysis (the 45 parameters tuples are tested to quantify parameters correlation). The parameters are strongly inter-dependent, the 45 tuples gather 87.3% of the variance, the parameters can not be calibrated separately. The weights of the parameter tuples confirm the idea of two parameters subsets: the 21 tuples of tissue specific parameters gather 49.6%, where the 21 tuples of tissue *v.s.* sample specific parameters gather only 35.6%, see Fig.3A, the 3 tuples of sample specific parameter are less correlated, they weigh only for 2.1%. The results of the first order are in Fig.3B and the details of the Sobol indices can be found in Appendix B.

The calibration process has been performed on all samples of the consolidation tests and on the mean curve of Test 1 of the indentation tests. It consists in a Newton algorithm on the 7 tissue specific parameters, the sample specific parameters being fixed. Once the algorithm gives no improvement (RMSE improvement below  $10^{-4}$ ), we perform, for each sample separately for the consolidation tests and for indentation Test 1, the calibration of the sample specific parameters  $E, k_{\text{int}}^s$  and  $S_0^l$ .

#### Measure of error

The error between numerical results  $y^{\text{num}}$  and experiments data  $y^{\text{exp}}$ , evaluated at  $n$  points, is measured in percentage by the relative root mean square error (RMSE):

$$RMSE = \sqrt{\sum_{i=1}^n \left( \frac{y_i^{\text{exp}} - y_i^{\text{num}}}{y_i^{\text{exp}}} \right)^2} \quad (12)$$

## 5. Results

The working hypothesis of this contribution is that the time-dependent properties of cortex tissue can be modeled using a two-phase flow poroelasticity framework. This hypothesis is

tested in this section to reproduce the experimental results of the consolidation tests in [14] and the unconfined indentation tests in [29]. To emphasise the limit of visco-elasticity compared to poro-elasticity, a 1D confined compression test with an available analytical solution is reproduced and discussed in Appendix A.

#### Consolidation tests

The mechanical response of the six samples is generated using parameters shown in Table 2. All the samples share the same *tissue specific* parameters set. Fig. 4A, B shows the results of the model for the 3 Newton and 6 Newton load series respectively. We obtain accurate results for the 6 samples with an error ranged from 1.1 to 5.6%. These results are obtained by calibrating three parameters, the Young's modulus  $E$ , the intrinsic permeability  $k_{\text{int}}^s$ , and the initial saturation of liquid phase  $S_0^l$  within the physiological range 6.5 – 17.2% (with one exception sample 3N C2 d at 5.9%). All the details are referenced in Table 2.

#### Indentation tests

The four indentation tests have been reproduced using the same mathematical modeling (section 3) and computational framework (section 4) than the consolidation tests. However, these indentation tests are not specifically adapted to poromechanical modeling, so that the nature of the free surface of the sample  $\Gamma_F$  (see Fig.2B) is not clearly prescribed from the fluids point of view. If the cortex slice cut follows a membrane, it leads to an impervious boundary as eq.13(a). In the case of a proper fluid drainage, this leads to a homogeneous Dirichlet condition, eq.13(b). The intermediate case leads to a semi-pervious boundary. To our knowledge, no investigation has been done on the passive drainage of the free surface for *ex vivo* cortex tissue testing, then we adopt a convective condition, usual in mass transfer through boundaries in poromechanics [49] eq.13(c).

$$\frac{\partial p^\alpha}{\partial n} = 0 \quad \text{on } \Gamma_F \quad \alpha = g, l \quad (13a)$$

$$p^\alpha = 0 \quad \text{on } \Gamma_F \quad \alpha = g, l \quad (13b)$$

$$\frac{\partial p^\alpha}{\partial n} = h(p^\alpha - p_0^\alpha) \quad \text{on } \Gamma_F \quad \alpha = g, l \quad (13c)$$

Our investigation shows that a semi-pervious boundary on the sample free surface gives slightly better results than an impervious boundary for the indenter surface response evaluated in this article. With the same parameters, the influence of different boundary conditions on the free surface are shown in Fig. 5. For the sake of simplicity, we choose an impervious boundary condition on the free surface.

We identify the parameters of [29] with only one test, the 1 millimeter diameter long range indentation on grey matter, denoted Test 1 in this article. Then, we validate these parameters on the other tests. The parameters of Test 1 are identified by the same process than consolidation tests, see section 4.

The resulting parameters of Test 1 (12.7% RMSE) are validated

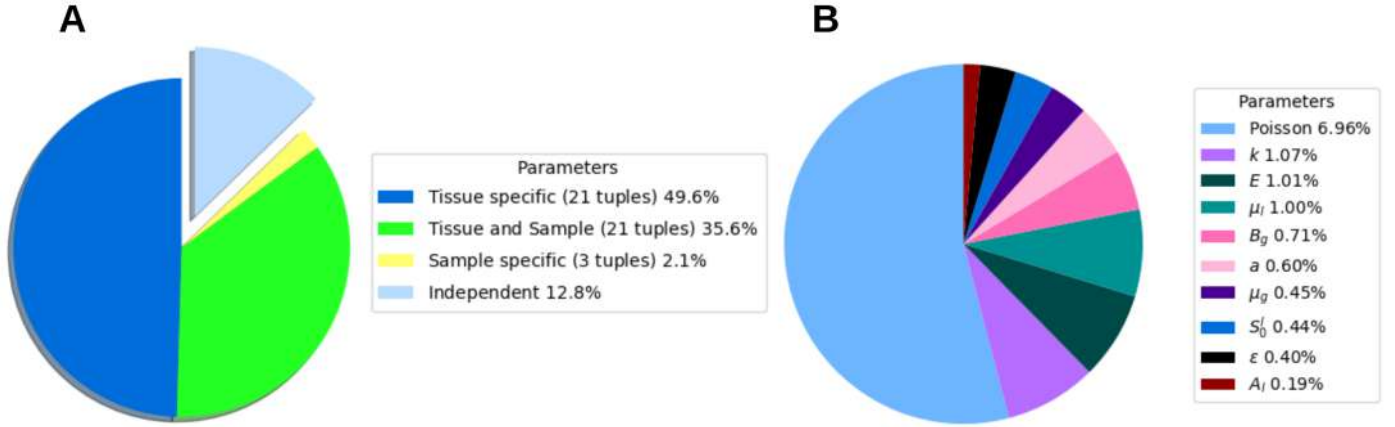


Figure 3: **Results of the second-order sensitivity analysis.** **A** The parameters interaction gathers 87.1% of the solution variance, *i.e.* the parameters are strongly correlated. The 21 tuples of tissue specific parameters weight for 49.6%, the 21 tuples of tissue/sample interaction weight for 35.6%, the 3 tuples of sample specific parameter are less correlated, they weight only for 2.1%. **B** Details of the weight of parameter independent sensitivity. The Poisson's ratio  $\nu$  sensitivity largely dominates the other parameters.

Table 2: **Model parameters calibrated of the consolidation tests on *ex vivo* human cortex [14].** Seven parameters,  $\varepsilon$ ,  $\nu$ ,  $\mu_l$  (Pa · s),  $A_l$ ,  $\mu_g$  (Pa · s),  $B_g$  and  $a$  (Pa) denoted *tissue specific*, are common to all samples. Three parameters,  $E$  (Pa),  $k_{int}^s$  (m<sup>2</sup>) and  $S_0^l$ , within the prescribed range [0.065; 0.232], are calibrated specifically for each sample. Only one sample, 3N C2 d, is slightly below  $S_0^l$  prescribed range.

Sample	$E$ (Pa)	$k_{int}^s$ (m <sup>2</sup> )	$S_0^l$	RMSE		
3N C2 b	4130	$2.2e^{-13}$	0.075	0.056		
3N C2 d	9300	$2.1e^{-13}$	0.059	0.036		
3N C2 e	3740	$4.7e^{-13}$	0.100	0.016		
6N C3 a	4330	$2.9e^{-13}$	0.114	0.011		
6N C3 b	3500	$6.1e^{-13}$	0.169	0.017		
6N 6	7100	$2.1e^{-13}$	0.094	0.015		
Shared parameters (tissue specific)		$S_0^l$ [0.065;0.232]	$\varepsilon$ 0.6 $\nu$ 0.49	$\mu_l$ (Pa · s) $8e^{-3}$	$A_l$ 1 $\mu_g$ (Pa · s) 35	$B_g$ 2 $a$ 800

on Test 2, 3 and 4, see Fig.6. It shows that the calibrated parameters lose accuracy but remain capable of reproducing the different tests. The change in the indenter diameter Test 2 slightly increases the error (13.5% RMSE). The results degradation is more significant when loading rates change in Test 3 (16.7% RMSE) or for the relaxation Test 4 (18.3% RMSE). Test 4 has been calibrated independently (10.3% RMSE), see Fig. 7, giving a different profile with  $E$  almost twice higher, see Table 3. The calibrated parameters of Test 1 give a value of  $S_0^l$  below the prescribed physiological range (< 6.5%), whereas the calibrated  $S_0^l$  of Test 4 is closer to the physiological range (see Table 3).

## 6. Discussion

The strong time-dependent mechanical behavior of the cortex is generally attributed to both the intrinsic viscoelasticity of the solid phase and fluid flow-induced poroelasticity but the relative contributions of the two are unclear. Conventional approaches combine viscoelastic and poroelastic behaviors into a single framework to be able to reproduce experimental data [14, 28].

In this contribution we propose to implement a two-phase flow poroelastic model of the cortex to capture the consolidation, the relaxation and the sensitivity of a set of selected load rates and investigate the dissipation mechanisms. Two sets of experiments are reproduced: the consolidation test of [14] and the unconfined indentation test of [29]. Our results show that the experimental curves can be reproduced without the need to resort to viscous effects, by adding an additional fluid phase.

First, we note that we examine in this paper two mechanical tests which are considerably different. For the consolidation test a load as high as 6 Newton is applied on the delicate structure of the cortex, which corresponds to a pressure of 8440Pa. In the indentation test, the load is approximately 0.6 mN (that is 4 orders of magnitude smaller), corresponding to a 20Pa pressure. The two tests are therefore very dissimilar in the way the sample is loaded, deformed and, possibly, damaged.

We estimate material parameters associated with the models we develop. We then compare those parameters with those published in the literature. We consider those parameters through the lens of the PhD thesis of Fanny Morin [50] (Chapter 6). Within this scope, we focus on rheological parameters extracted from confined/unconfined experiments on



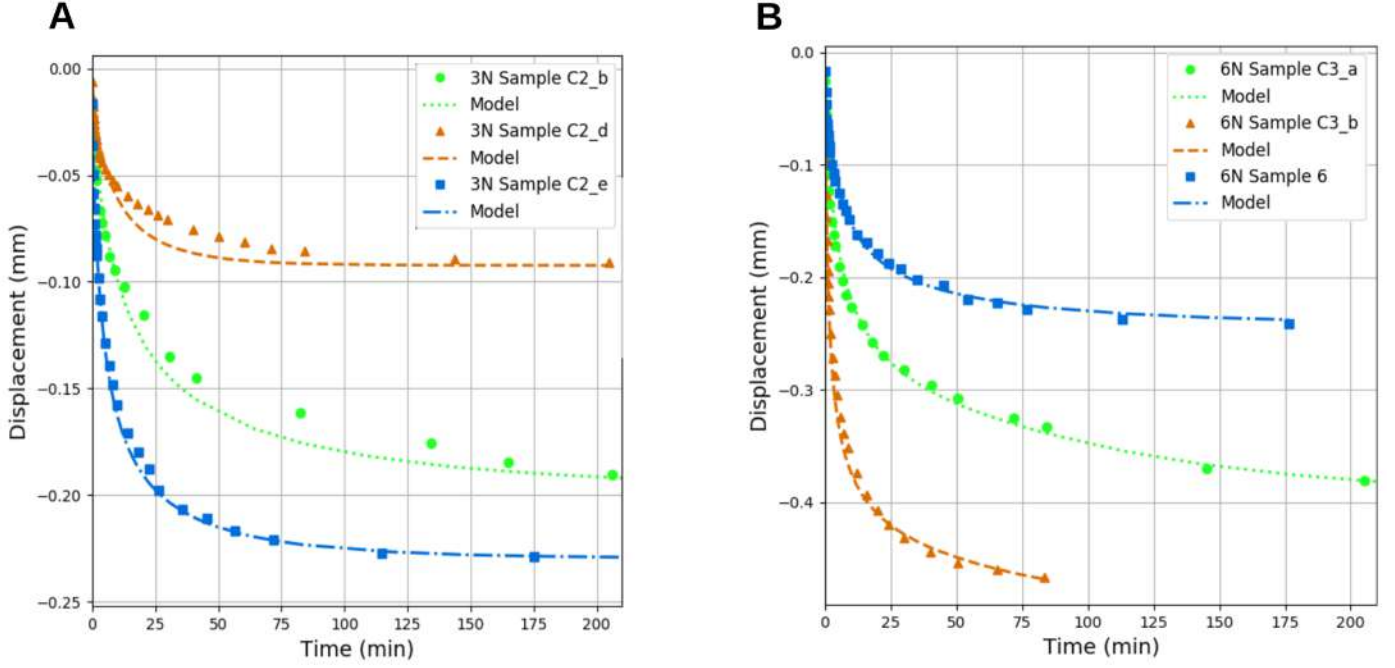


Figure 4: **Numerical reproduction of Franceschini *et al.* consolidation tests on *ex vivo* human cortex [14].** Time (minutes) vs. Displacement (mm). **A** 3 Newton load series. Sample 3N C2 b (experimental green circle; numerical green dotted) is reproduced with 0.056 RMSE (see Eq.12); Sample 3N C2 d (experimental orange triangle; numerical orange dotted) with 0.036 RMSE; Sample 3N C2 e (experimental blue square; numerical blue dotted) with 0.016 RMSE. **B** 6 Newton load series. Sample 6N C3 a (experimental green circle; numerical green dotted) is reproduced with 0.011 RMSE; Sample 6N C3 b (experimental orange triangle; numerical orange dotted) with 0.017 RMSE; Sample 6N 6 (experimental blue square; numerical blue dotted) with 0.015 RMSE.

Table 3: **Parameters for indentation tests on *ex vivo* bovin cortex.** Validation (V) on Test 2, 3 and 4 of the calibrated (C) parameters of the Test 1. Specific calibration (C) of the Test 4. Seven parameters,  $\varepsilon$ ,  $\nu$ ,  $\mu_l$  (Pa · s),  $A_l$ ,  $\mu_g$  (Pa · s),  $B_g$  and  $a$  (Pa) denoted *tissue specific*, are common to all samples.

Sample	$E$ (Pa)	$k_{int}^s$ (m <sup>2</sup> )	$Sl_0$	RMSE					
Test 1 (C)	605	$2.8e^{-12}$	0.027	0.127					
Test 2 (V)	605	$2.8e^{-12}$	0.027	0.135					
Test 3 (V)	605	$2.8e^{-12}$	0.027	0.167					
Test 4 (V)	605	$2.8e^{-12}$	0.027	0.183					
Test 4 (C)	1100	$4.2e^{-12}$	0.047	0.099					
Shared parameters (tissue specific)		$Sl_0$ [0.065;0.232]	$\varepsilon$ 0.5	$\nu$ 0.47	$\mu_l$ (Pa · s) $3^{-3}$	$A_l$	$\mu_g$ (Pa · s) 1	$B_g$ 30	$a$ 1 400

human/animal grey matter.

The stiffness values (Young’s modulus) reported in the literature, computed by considering grey matter as a monolithic solid, are widely scattered (1.1 to 8 kPa). In the unconfined indentation case, these stiffness values are much higher than those we obtain (0.6-1.2kPa). This observation is explained by the fact that the stiffness we “measure” consists in both a solid component and a fluid component, thereby decreasing the effective stiffness contribution of the solid phase Young’s modulus. The values we obtain for  $E$  in the confined consolidation tests are six times as high as those obtained in the unconfined indentation case ( $E = 5.35 \pm 2.12$  kPa). This could seem surprising. The possibility of different stiffness between human and animal - simian, bovine and porcine - brain have led to contradictory results (see [51] and [52]). We looked for a possible explanation for this stiffness discrepancy within environmental conditions such as: sample preservation, hydra-

tion and temperature. The loss of hydration and temperature have cumulative effects. All other conditions remaining equal, temperatures of 24°C and 37°C will lead to a stiffness 40% lower, but the same variation with dehydration will lead to a stiffness 5 fold higher (see Forte *et al.* [53] for a detailed study on this topic). However, according to the ranges given in [53], the confined consolidation and unconfined indentation tests were made in sufficiently close environmental conditions to rule out this hypothesis.

We therefore hypothesise that, during confined consolidation, pore locking may take place, leading to the creation of fluid “pockets” within the structure. This incomplete consolidation results in an effective stiffening of the sample and thereby in an increase of the apparent Young’s modulus. Regarding the Poisson’s values, they correspond to the literature consensus with  $\nu = 0.47$  in the unconfined case, and

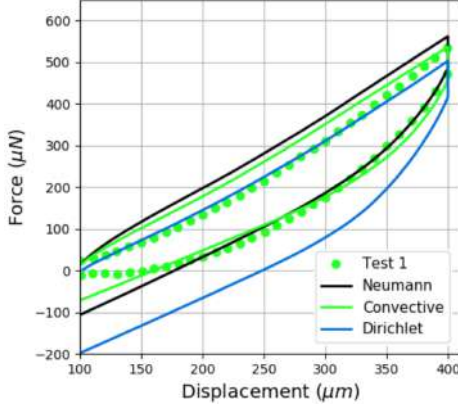


Figure 5: **Influence of the free surface boundary condition on the indenter surface response.** Shared parameter set:  $E = 730 \text{ Pa}$ ,  $k_{\text{int}}^s = 3 \cdot 10^{-3} \text{ m}^2$ ,  $S_0^l = 0.063$ ,  $\varepsilon = 0.6$ ,  $\nu = 0.49$ ,  $\mu_l = 3 \cdot 10^{-3} \text{ Pa} \cdot \text{s}$ ,  $A_l = 1$ ,  $\mu_g = 30 \text{ Pa} \cdot \text{s}$  and  $B_g = 2$ . Experimental (green dotted); Neumann (black) impervious condition eq.13(a); Dirichlet (blue) drained condition eq.13(b); Convective (green), with the additional parameter  $h = 10^{-4}$ , eq.13(c) semi-pervious condition.

$\nu = 0.49$  in the confined case.

In the confined case, we obtain one order of magnitude smaller permeabilities than in the unconfined case. The intrinsic permeabilities we evaluate, amended by their respective relative permeabilities, are however within the experimental range provided in the literature. Such decrease in permeability can be caused by the pore locking phenomenon described above. Moreover, the consolidation tests were performed within 12 hours, as opposed to 6 hours for the indentation tests. This time delay could lead to significant changes in the microstructure [48], which also alters the permeability.

The review of Budday *et al.* [7] pointed out the promising coupling of poro-viscoelasticity, as it takes account of both the interplay between interstitial fluid and stroma and the time-dependent response of cellular phase. The model proposed by Franceschini *et al.* in [14] along with their experiments is an example of this coupling. We show in this article that a one-phase flow coupled with a viscoelastic solid, in the reproduced loading scenarii, can be replaced by a two-phase flow with an elastic solid. The two different dynamic viscosities ( $\mu_l = 5.5 \pm 2.5 \cdot 10^{-3} \text{ Pa} \cdot \text{s}$  and  $\mu_c = 32.5 \pm 2.5 \text{ Pa} \cdot \text{s}$ ) give two different characteristic times. They are comparable to the two-terms Prony series proposed in the viscoelastic parameter identification of human brain tissue of Budday *et al.* in [11] characterised by  $\tau_1 = 0.18 \text{ s}$  and  $\tau_2 = 63.5 \text{ s}$ . Moreover, our parameters are not dependent on the geometry, as shown in the Appendix A. The fact that the tissue characteristics are build upon experimental findings offers a reliable way for a tissue specific pre-calibration. The sample calibration is only done on three parameters, which is equivalent, in computational cost, to the classic hyperelastic models (Yeoh [54], Ogden [55]).

Even if we endeavour to build our model on experimental matter, we are aware of its limits. A solid scaffold described

as the sum of the stroma, the vascular network and the ECM is obviously non-linear. We have not included dependency between permeability and porosity in our model (the interested reader could find a presentation in [56]), whereas in confined deformation with an incompressibility hypothesis, the main part of the deformation is absorbed by the pores. Adding this relation would lead to closer values of intrinsic permeability  $k_{\text{int}}^s$  between indentation and consolidation, and partially suppress the artificial stiffness of consolidation parameters, by increasing the fluids supported stress. We can also point out that the total stress relationship we used, along with a Biot coefficient equal to 1 is a strong hypothesis. The incompressibility of the fluids is not to be questioned. Yet, micro-structural changes due to post-mortem experiments or mechanical damage may increase solid scaffold compressibility. Only three load rates are reproduced in this paper ( $1 \mu\text{m/s}$ ,  $5 \mu\text{m/s}$  and  $100 \mu\text{m/s}$ ), a large panel still remains to explore. Likewise, our model response on various loading scenarii - tension or shear - or experimental condition - such as temperature dependency - remains open. As a consequence, our results are only relevant at room temperature under humidity control.

The results in this paper indicate that multi-phase models could be a strong basis for the description of biological tissue such as the brain, *ex vivo* and *in vivo*, as some advanced models can reproduced interplay of vessels and tissue [57, 30]. We hope this contribution will encourage the community to develop new experimental techniques which are compatible with poro-mechanical models. More specifically, the enforcement of boundary conditions on free surfaces could help the modeling of different drainage conditions.

Beyond experimental setups, another open challenge lies in the ability of models to be transferred from one observational/experimental situation to another. For this work to have impact in a clinical setting, methods must be developed to transfer parameters and models from one set of patients to another. We believe that a quantitative approach to this would be to develop robust model selection approaches from experimental or clinical data [58].

## Acknowledgments

The results presented in this paper were carried out using the HPC facilities of the University of Luxembourg [59] (see <https://hpc.uni.lu>).

## Appendix A. Viscoelastic modeling limitation on consolidation test reproduction

Let us consider two reference models: a one solid-one fluid poroelastic model and a rheological viscoelastic model constituted by a Kelvin-Voigt chain; and use these two models to simulate a 1D confined compression test. In this test the tissue is constrained in a cylindrical chamber and compressed at the top surface with a constant pressure  $P_0$ . The specimen is

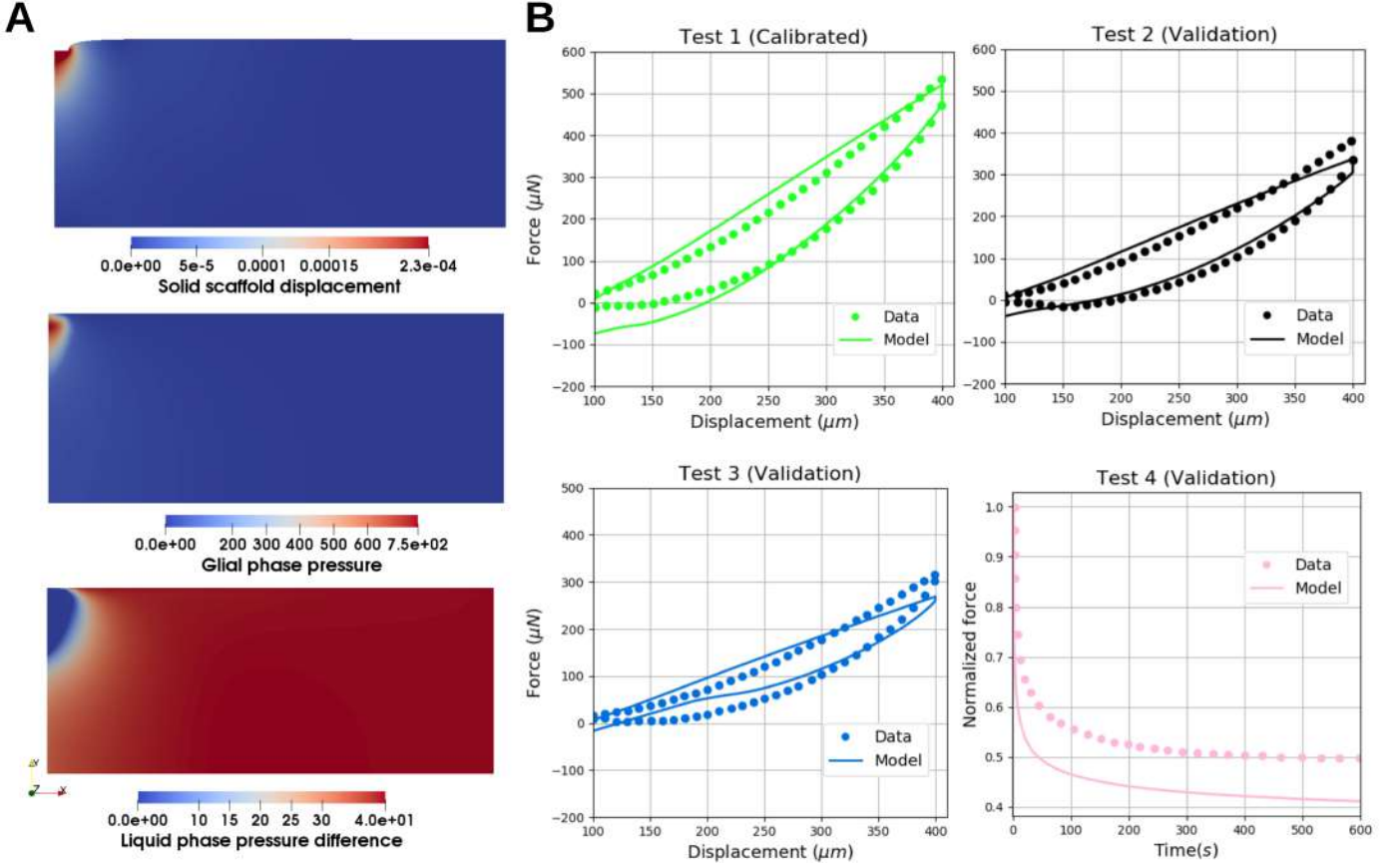


Figure 6: **Indentation tests on *ex vivo* bovine cortex [29]; validation of the calibrated parameters on Test 1.** **A** Example of the simulation results on Test 1. At  $230\ \mu\text{m}$  indentation depth, the glial pressure raised to  $780\ \text{Pa}$  and flush out the liquid phase (*i.e.*  $0\ \text{Pa}$  of pressure difference  $p^{ls}$  in this area). Note that the liquid phase is subjected to a pressure  $p^l$  of a few hundreds of Pascal at the indenter zone and only  $40\ \text{Pa}$  at the sample boundaries. **B** Test 1, experimental (green dotted), numerical (green plain line), RMSE= 12.7%; Test 2, experimental (black dotted), numerical (black plain line), RMSE= 13.5%; Test 3, experimental (blue dotted), numerical (blue plain line), RMSE= 16.7%; Test 4, experimental (pink dotted), numerical (pink plain line), RMSE= 18.3%.

fully sealed with the exception of the top and the bottom surface where a porous membrane allows drainage of the inside fluid during the test. The geometry and boundary conditions of the test are represented in Fig. 4A. The simplicity of the two considered models jointed with the simplicity of the considered test allows to derive the analytical solution for both modeling approaches (see [60] for the poromechanical model). For the one-fluid one-solid poroelastic model:

$$p_z(z, t) = P_0 \frac{4}{\pi} \sum_{k=1}^{\infty} \frac{(-1)^{k-1}}{2k-1} \cos\left((2k-1)\frac{\pi z}{2L}\right) \exp\left((2k-1)^2 \frac{\pi^2 c_v t}{4h^2}\right) \quad (\text{A.1})$$

with the consolidation coefficient under the hypothesis of phases incompressibility:

$$c_v = \frac{kM}{\mu} \quad (\text{A.2})$$

with  $M$  the longitudinal modulus:

$$M = \frac{E(1-\nu)}{(1+\nu)(1-2\nu)} \quad (\text{A.3})$$

$E$  the Young's modulus and  $\nu$  the Poisson's ratio.

For the viscoelastic Kelvin-Voigt chain:

$$u_z(z, t) = \frac{P_0 z}{M} \left[ 1 - \exp\left(-\frac{Et}{\eta}\right) \right] \quad (\text{A.4})$$

We see that the poroelastic model is governed by the parameters  $E$ ,  $\nu$  and the ratio  $\frac{k}{\mu}$  and the viscoelastic model by  $E$ ,  $\nu$  and  $\eta$ . However, the analytical solution of the porous model contains  $h$ , the sample height, which strongly influence the consolidation time. Then, once the parameters of the porous model are calibrated, they will remain relevant when the height of the sample varies.

## Appendix B. Variance-based local sensitivity analysis

We performed a variance-based local sensitivity study of the finite element solution on the parameters as follows:

- A first-order analysis, the 9 parameters are disturbed one at a time respectively of  $\pm 10\%$ .
- Interaction analysis, the 36 parameters tuples are evaluated simultaneously disturbed.

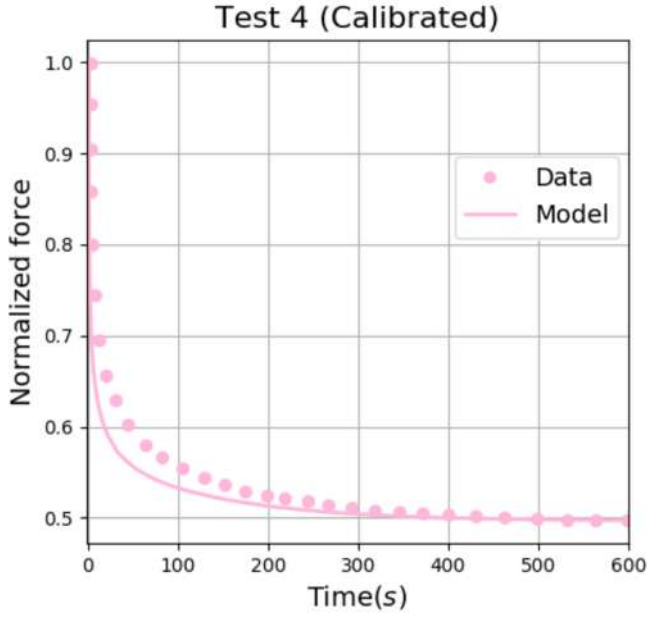


Figure 7: **Indentation tests on *ex vivo* bovine cortex [29]: calibration of the parameters of Test 4.** Experimental (pink dotted), numerical (pink plain line), RMSE= 10.3%.

All the results were interpreted with a polynomial model in order to quantify their weights in the solution variance, referred to as Sobol indices. The initial guess of the parameters set was:  $E = 3500$  Pa,  $\nu = 0.45$ ,  $k = 10^{-13}$  m<sup>2</sup>,  $\varepsilon = 0.55$ ,  $\mu_l = 8 \cdot 10^{-3}$  Pa·s,  $\mu_g = 35$  Pa·s,  $A_l = 1$ ,  $B_g = 2$  and  $S_0^l = 0.012$ . It gives the base error  $J_0$ .

#### First-order analysis

Each parameter is disturbed one at a time respectively of  $\pm 10\%$ , giving the corresponding error  $\tilde{J}$ . The relative variations of the error were calculated as follows:

$$\text{Var} = \frac{\tilde{J} - J_0}{J_0} \quad (\text{B.1})$$

where  $J_0$  is the error with the parameters at their initial values. In order to quantify the impact of each parameter, the following linear model was set:

$$\text{Var} = 1 + \sum_i \theta_i \alpha_i \quad (\text{B.2})$$

where  $\alpha_i$  is an auxiliary parameter  $\in [-1, +1]$  representing the perturbations of  $\pm 10\%$  of the  $i^{\text{th}}$  parameter and  $\theta_i$  the slope of the variation.

In a first-order analysis, the influence of the  $i^{\text{th}}$  parameter is given by the Sobol indices:

$$S_i = \frac{\theta_i^2}{\sum_i \theta_i^2} \quad (\text{B.3})$$

The results of the first-order analysis are reported in Table B.4.

Table B.4: **Sobol indices of the first-order local sensitivity analysis.**

Parameter	$\theta$	$S_i(\%)$
$E$	0.0426	8.29
$\nu$	0.2384	56.7
$k_{\text{int}}^s$	-0.0430	8.74
$\varepsilon$	-0.0304	3.32
$\mu_l$	0.0346	8.17
$\mu_g$	0.0114	3.70
$A_l$	0.0002	1.58
$B_g$	0.0208	5.80
$S_0^l$	-0.0256	3.63

#### Interaction analysis

We evaluate the correlation between parameters. The 36 tuples  $(\alpha_i, \alpha_j)_{i>j}$  have been evaluated at  $\pm 10\%$ . The corresponding polynomial model becomes:

$$\text{Var} = 1 + \sum_i \theta_i \alpha_i + \sum_{ij, i>j} \theta_{ij} \alpha_i \alpha_j \quad (\text{B.4})$$

with the respective Sobol indices:

$$S_i = \frac{\theta_i^2}{\sum_i \theta_i^2 + \sum_{ij, i>j} \theta_{ij}^2} \quad \text{and} \quad S_{ij} = \frac{\theta_{ij}^2}{\sum_i \theta_i^2 + \sum_{ij, i>j} \theta_{ij}^2} \quad (\text{B.5})$$

The results are reported in Table B.5.

Table B.5: Sobol indices of the second-order local sensitivity analysis.

Parameters correlation			Parameters independence			
Tissue specific 49.5%	$S_{ij}(\%)$	Tissue/Sample 35.4%	Sample specific 2.1%	$S_{ij}(\%)$	$S_i(\%)$	
$(\nu, \varepsilon)$	5.34	$(E, \nu)$	$(E, k)$	0.45	$E$	12.8%
$(\nu, \mu_l)$	7.59	$(E, \varepsilon)$	$(E, S_0^l)$	0.26	$\nu$	6.96
$(\nu, \mu_g)$	6.59	$(E, \mu_l)$	$(k_{int}^s, S_0^l)$	1.36	$k_{int}^s$	1.07
$(\nu, A_l)$	6.19	$(E, \mu_g)$			$\varepsilon$	0.40
$(\nu, B_g)$	7.14	$(E, A_l)$			$\mu_l$	1.00
$(\nu, a)$	7.06	$(E, a)$			$a$	0.60
$(\varepsilon, \mu_l)$	0.31	$(E, B_g)$			$\mu_g$	0.45
$(\varepsilon, \mu_g)$	0.33	$(k_{int}^s, \nu)$			$A_l$	0.19
$(\varepsilon, A_l)$	0.37	$(k_{int}^s, \varepsilon)$			$B_g$	0.71
$(\varepsilon, B_g)$	0.37	$(k_{int}^s, \mu_l)$			$S_0^l$	0.44
$(\varepsilon, a)$	0.43	$(k_{int}^s, a)$				
$(\mu_l, \mu_g)$	1.16	$(k_{int}^s, \mu_g)$				
$(\mu_l, A_l)$	0.88	$(k_{int}^s, A_l)$				
$(\mu_l, B_g)$	1.38	$(k_{int}^s, B_g)$				
$(\mu_l, a)$	1.37	$(S_0^l, \nu)$				
$(\mu_g, A_l)$	0.30	$(S_0^l, \varepsilon)$				
$(\mu_g, B_g)$	0.81	$(S_0^l, \mu_l)$				
$(\mu_g, a)$	0.77	$(S_0^l, a)$				
$(A_l, B_g)$	1.14	$(S_0^l, \mu_g)$				
$(A_l, a)$	0.49	$(S_0^l, A_l)$				
$(B_g, a)$	1.40	$(S_0^l, B_g)$				

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